Anti-inflammatory effects of resveratrol in lung epithelial cells: molecular mechanisms


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Resveratrol (3,4’,5-trihydroxystilbene) is a polyphenolic stilbene found in the skins of red fruits (22) (Fig. 1). It is thought that the presence of resveratrol in red wine may, in part, explain the “French paradox” and be responsible for many of the health benefits ascribed to consumption of red wine (21, 30, 47). Indeed, the related stilbene quercetin exhibits antioxidant and antiproliferative capacities similar to those of resveratrol (17, 30, 47).

More recently, the anti-inflammatory effects of resveratrol have been associated with inhibition of the transcription factor NF-κB (34), possibly mediated via the inhibition of IκB kinase (24). NF-κB activation is required for the expression of many inflammatory proteins, such as granulocyte-macrophage colony-stimulating factor (GM-CSF), IL-8, COX-2, and inducible nitric oxide synthase (iNOS) (4, 39). Therefore, inhibition of NF-κB could reduce the expression of inflammatory genes and is one mechanism by which the anti-inflammatory agents glucocorticosteroids might elicit their anti-inflammatory effects (38). A second transcription factor, activator protein-1 (AP-1), may also be inhibited by resveratrol (34). NF-κB and AP-1 may be important in the regulation of many genes that are induced via oxidative stress (29) and might in part explain some of the antioxidative properties of resveratrol. This suggests that an understanding of the mechanism(s) of action of these compounds may lead to novel anti-inflammatory therapies.

The mainstays of anti-inflammatory therapy in inflammatory airway diseases, including asthma, are the glucocorticosteroids. However, there is a small but significant group of asthmatic patients for whom glucocorticosteroids are not effective: either high doses are needed that have systemic side effects, or very rarely patients are completely resistant (31). In contrast, glucocorticosteroids are ineffective at suppressing the
airway inflammation of chronic obstructive pulmonary disease (COPD) (11). Recently, we demonstrated that resveratrol inhibited IL-8 and GM-CSF release by alveolar macrophages from patients with COPD (12) under conditions where steroids are largely ineffective (13). Therefore, this study aimed to investigate the mechanism of action of plant-derived polyphenolic compounds, such as resveratrol and quercetin, by examining inflammatory mediator release by human lung epithelial cells.

MATERIALS AND METHODS

Materials

A549 cell lines were obtained from the American Type Culture Collection. DMEM, Ham’s F-12 nutrient medium, HBSS, and G-418 were purchased from GIBCO (Paisley, Scotland); 2,3-diaminonaphthalene from Alexis (Nottingham, UK); IL-1β, TNF-α, IFN-γ, and IL-8 ELISA kits from R & D Systems Europe (Abingdon, Oxon, UK); anti-GM-CSF antibodies from PharMingen (San Diego, CA); avian myeloblastosis virus reverse transcriptase, RNAsin, random primers, Tfx50 reagent, restriction enzymes, and pGL.neo.TATA from Promega (Southampton, UK); pMC1neoPolyA from Stratagene (La Jolla, CA); Taq polymerase, dNTPs, and KCl buffer from Bioline (London, UK); RNA extraction kit from Qiagen (Crawley, UK); WY-14643 from Tocris Avonmouth (Avon, UK). Resveratrol was a kind gift from Pharmascience (Montreal, PQ, Canada). GW-466471 was a kind gift from GlaxoSmithKline (Stevenage, Herts, UK). Sigma Chemical (Poole, UK) or BDH (Poole, UK) supplied all other reagents.

Methods

Isolation and culture of human primary epithelial cells. Human primary epithelial cells were obtained from explant cultures of tissue obtained from normal human lung transplant donors as described previously (15). Cells were grown to 90% confluence and cultured in serum-free medium for 24 h before initiation of experiments. All experiments were performed under serum-free conditions. Ethical permission for this study was obtained from the Ethics Committee of the Royal Brompton Hospital.

Culture of A549 cell lines. A549 cells were cultured in DMEM containing 10% (vol/vol) FCS and 2 mM glutamine. Cells were maintained in a humidified incubator at 37°C and 95% (vol/vol) air-5% (vol/vol) CO2. The cells were provided with fresh medium every 2–3 days. The cells were grown to 90% confluence and cultured in serum-free medium for 24 h before initiation of experiments. All experiments were performed under serum-free conditions.

Measurement of cell viability. Cells were treated with 1 mg/ml 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) in HBSS for 30 min at 37°C. The MTT solution was removed from the surface of the cells, and DMSO was added. The absorbance of the resultant solution was measured at 550 nm, and treated cells were compared with control cells.

Measurement of nitrite and nitrate in cell culture medium. The amount of nitrite and nitrate in cell culture medium was measured using a modification of the method of Misko et al. (37) as described previously (15).

Measurement of GM-CSF and IL-8 in cell culture medium. GM-CSF was measured by sandwich ELISA, as described previously, with a detection limit of 32 pg/ml (15). IL-8 was measured using a commercially available ELISA kit (R & D Systems), according to the manufacturer’s instructions, with a detection limit of 32 pg/ml.

Measurement of PGE2 release from A549 cells. PGE2 release was measured using RIA as described previously (7).

Measurement of histone deacetylation and histone acetylation activity. Histone deacetylation activity and histone acetylation activity were measured in A549 cells as described previously by Ito et al. (27).

Generation of stable A549 reporter plasmids. A549 cells harboring stable transfectants of the NF-κB-dependent reporter 6xBtkluc.neo
have been described previously (3). An identical protocol was used to generate stable transfectants containing six cAMP regulatory elements (CRE) or six AP-1 sites driving a luciferase gene (23). The glucocorticoid regulatory element (GRE) reporter was constructed from pGL3basic.neo. This vector was then opened at the SalI/HindIII site, containing a C32T mutation at position −31 to generate a consensus TATA box was inserted into the HindIII site of pGL3.neo.TATA. This vector was then opened at the SalI site, and a double-stranded oligonucleotide, 5′-HinTATA box was inserted into the containing a C32T mutation at position −31 to generate a consensus TATA box was inserted into the HindIII site of pGL3.neo.TATA. This vector was then opened at the SalI site, and a double-stranded oligonucleotide, 5′-GCT GTA CAG GAT GTT CTA GGC TGT ACA GGA TGT TCT AG-3′, containing two consensus GRE sites (underlined) was inserted to create pGL3.neo.TATA. Stable recombinants in A549 cells were generated as described previously (7). In all cases, before treatment, cells were incubated overnight in serum-free medium in the absence of G-418 selection.

Measurement of luciferase activity. Luciferase activity was measured using the luciferase assay system (Promega). The cell lysates were resuspended in 200 μl of cell lysis buffer, and 40 μl of the resulting lysate were used to measure luciferase activity in the presence of 40 μl of luciferase assay reagent. Emitted light was measured using a luminometer (Turner Designs model TD-20/20, Steptech Instruments, Stevenage, UK).

RNA isolation. RNA was isolated from primary epithelial cells using the Qiagen RNeasy mini kit according to the manufacturer’s instructions.

RT-PCR. Reverse transcription was performed on 0.5 μg of RNA. RNA was heated to 70°C for 5 min and then mixed with 0.01 μg/μl random primers, 1.0 mM dNTP, 1 U/μl RNAseIn, and 0.25 U/μl avian myeloblastosis virus reverse transcriptase in 1× reverse transcriptase buffer and incubated at 42°C for 1 h and then denatured at 90°C for 4 min. The resultant cDNA was diluted by the addition of 80 μl of water.

For PCR, 5 μl of cDNA were incubated in a final volume of 25 μl containing 1× KCl buffer, 2 mM dNTP, 5 ng/μl specific primers, and 0.02 μl/μl Taq polymerase. PCR for iNOS, COX-2, and GAPDH was performed as previously reported (16, 40).

Specific primers for tPA gave a specific product of 531 bp: 5′-CCCTGGCAGGCTGCCATGT-3′ (forward primer) and 5′-GTTCTTCCAGCACACAGCATG-3′ (reverse primer). The cycles were as follows: 94°C for 45 s, 56°C for 60 s for 32 cycles followed by 72°C for 10 min.

PCR products were identified on 2% (wt/vol) agarose gels. Samples that did not contain reverse transcriptase were used as negative controls.

RESULTS

Effect of Resveratrol and Related Compounds on Inflammatory Mediator Release From A549 Cells

To investigate the mechanism of resveratrol inhibition of inflammatory mediator release, the effect of this drug and the related compounds quercetin and deoxyrhapontin was examined using A549 cells. IL-1β (1 ng/ml) stimulated these cells to release GM-CSF: 35.7 ± 1.47 and 741 ± 270.5 pg/ml in basal and stimulated conditions, respectively (n = 9). Resveratrol inhibited the release of GM-CSF from IL-1β-stimulated A549 cells by ∼75%, with an IC50 of 24.3 ± 5.5 μM (n = 9; Fig. 2A). Quercetin also inhibited GM-CSF release from these cells, with less efficacy than resveratrol but with similar potency (∼50%), with an IC50 of 21.6 ± 12.3 μM (n = 8; Fig. 2B). However, deoxyrhapontin failed to inhibit GM-CSF release from A549 cells (Fig. 2C). Resveratrol at 100 μM could inhibit IL-8 release from A549 cells that had been stimulated with 1 ng/ml IL-1β (undetectable and 15.4 ± 3.8 ng/ml in basal and stimulated conditions, respectively, n = 6) by only ∼40% (IC50 = 72 ± 11 μM, n = 6; Fig. 3A). Quercetin inhibited IL-8 release from A549 cells to a similar level, with an EC50 of 48 ± 24 μM (n = 6; Fig. 3B), whereas deoxyrhapontin had no inhibitory effect (Fig. 3C). Cell viability as determined by MTT assays was unaffected by any of these treatments.

Effect of Estrogens

To investigate whether resveratrol-mediated inhibition of inflammatory mediator release from human airway epithelial cells was via the estrogen receptor, the effects of the estrogen antagonist tamoxifen on resveratrol-mediated inhibition were examined. Tamoxifen at 1–20 μM has been demonstrated by other workers (9) to affect arachidonic acid release from A549 cells. Tamoxifen (10 μM) failed to antagonize the resveratrol-mediated inhibition of GM-CSF or IL-8 release from IL-1β-stimulated A549 cells (Fig. 4, A and B). IL-1β (1 ng/ml) stimulated these cells to release PGE2 (0.13 ± 0.05 and 4.07 ±
1.26 pg/ml in basal and stimulated conditions, respectively, n = 4). Similarly, resveratrol inhibition of IL-1β-stimulated PGE2 release (IC50 = 16.1 ± 3.0 μM) could not be antagonized by tamoxifen (Fig. 4C). Indeed, tamoxifen further suppressed the release of PGE2 from these cells. In addition, estradiol was unable to inhibit GM-CSF, IL-8, or PGE2 release from these cells (Fig. 4), suggesting that the observed inhibitory effects of resveratrol are not mediated via the estrogen receptor.

Effect of Glucocorticoids

Structural similarities between resveratrol and glucocorticosteroids suggested that resveratrol could be acting as a glucocorticosteroid. Mifepristone is a glucocorticosteroid receptor antagonist that exhibits little ability to transactivate glucocorticosteroid receptor-dependent transcription (46) and has a K_i for dexamethasone binding in human airway epithelial cells of ~60 nM (32). To investigate the possibility that resveratrol is acting via the glucocorticosteroid receptor, the effect of the glucocorticosteroid antagonist mifepristone (RU-486) was examined. Mifepristone at 10^-8 – 10^-6 M has been shown to antagonize dexamethasone-mediated repression of COX-2 expression in A549 cells (41). Mifepristone was unable to antagonize resveratrol-mediated inhibition of IL-1β-stimulated IL-8 release from A549 cells (Fig. 5A). This suggested that resveratrol was not exerting its effects via the glucocorticoid receptor. To confirm this observation, A549 cells were stably transfected with a luciferase reporter gene containing 2XGRE sites in front of a luciferase reporter gene. This reporter was activated by dexamethasone (Fig. 5B). By contrast, increasing concentrations of resveratrol failed to activate the luciferase reporter (Fig. 5B). This would suggest that resveratrol is not acting as an agonist for the glucocorticoid receptor.

Effect on Histone Acetylation

To further characterize the mechanism of resveratrol action, its effect on the regulation of gene expression was investigated. One such possible mechanism is the unwinding of DNA from around histone proteins, thereby allowing the binding of transcription factors. This is regulated via acetylation of histones by histone acetylase transferases (HAT). Acetylation of histones alters their confirmation, such that DNA unwinds, allowing transcription factors to bind DNA, enabling gene transcription to occur. This regulation can be silenced by the removal of acetyl moieties from histones by histone deacetylases (HDAC). Hence, gene transcription may be regulated through the acti-
Fig. 5. Effect of resveratrol on glucocorticoid activity. A: A549 cells were treated for 24 h in the absence or presence of 1 ng/ml IL-1β in the absence or presence of 50 μM resveratrol (Res) with or without 0.01–1 μM mifepristone (Mif). Medium was harvested, and IL-8 levels were measured by ELISA. B: A549 cells that had been stably transfected with 2XGRE luciferase reporter gene were treated for 6 h with increasing concentrations of dexamethasone (●) or resveratrol (○), cells were harvested, and luciferase activity was measured.

Effect on Transcription

A luciferase reporter gene containing six kB sites was stably transduced into A549 cells, and the effect of resveratrol and quercetin on transcription factor activation was investigated. Resveratrol inhibited IL-1β-induced expression of luciferase in a dose-dependent manner (Fig. 6A), with an IC₅₀ of 41 ± 10 μM (n = 7). Moreover, complete inhibition of this reporter gene activity occurred in the presence of 100 μM resveratrol (Table 1). Quercetin also inhibited IL-1β-induced expression of luciferase (IC₅₀ = 21 ± 13 μM, n = 5; data not shown). Although dexamethasone inhibited expression of the NF-kB reporter, the maximum inhibition observed was only 41% (IC₅₀ = 10 ± 6 nM, n = 5; Fig. 6A, Table 1). The effect of these compounds was also examined using an AP-1 luciferase reporter. Cells were stimulated with 10⁻⁷ M PMA in the presence of the test compounds. Again, resveratrol and quercetin inhibited AP-1-dependent luciferase expression [IC₅₀ = 28 ± 13 μM (Fig. 6B) and 20 ± 12 μM, n = 4–6 (data not shown)]. Dexamethasone inhibited AP-1-dependent luciferase expression, but again maximum inhibition was less than that seen with resveratrol or quercetin (Fig. 6B, Table 2). Another transcription site implicated in the regulation of inflammatory genes is CRE. Again, a luciferase reporter was generated in A549 cells, and activity was stimulated with IL-1β. Resveratrol and quercetin inhibited CRE-dependent expression, with IC₅₀ values of 46 ± 23 μM (Fig. 6C) and 49 ± 28 μM (n = 6; data not shown). Again dexamethasone inhibited CRE-dependent transcription, but with a maximum inhibition of ~50% in the presence of 100 μM dexamethasone (Fig. 6C, Table 2). These results would suggest that the plant-derived compounds resveratrol and quercetin inhibit inflammatory gene transcription and, although not as potent as dexamethasone, inhibit transcription factor activity to a greater extent than the steroid used in these studies.

To ensure that the inhibitory effect of resveratrol was not merely occurring through a general inhibition of gene transcription, the effect of resveratrol was examined on a luciferase reporter gene containing a TATA transcription site. Resveratrol failed to inhibit constitutive luciferase expression in this system (data not shown), suggesting that the inhibitory effect of resveratrol on inflammatory gene transcription is not merely a reflection of inhibition of gene transcription per se.

Effect of PPAR-α

The possibility that the anti-inflammatory action of resveratrol could be via activation of the PPAR-α receptor was examined using the PPAR-α agonist WY-14643 and the specific PPAR-α antagonist GW-466471 (48). The PPAR-α agonist WY-14643 was less effective than resveratrol at inhibiting IL-1β-dependent NF-κB transcription (EC₅₀ = 57.4 ± 21.5 μM, n = 3; Fig. 7A). Moreover, the specific PPAR-α antagonist GW-466471 was unable to reverse the resveratrol-mediated inhibition of IL-1β-dependent NF-κB transcription (Fig. 7B). Taken together, these data suggest that resveratrol is not mediating its anti-inflammatory effects in A549 cells via PPAR-α.

Effect of Resveratrol on Inflammatory Mediator Release From Primary Human Airway Epithelial Cells

Human primary epithelial cells were cultured in the presence of 50 ng/ml of IL-1β, TNF-α, and IFN-γ (“cytomix”) in the

Table 1. Effect of resveratrol and quercetin on HDAC and HAT activity in A549 cells

<table>
<thead>
<tr>
<th>HDAC Activity, dpm/μg protein</th>
<th>HAT Activity, dpm/10 μg protein</th>
</tr>
</thead>
<tbody>
<tr>
<td>No stimulus</td>
<td>206±28</td>
</tr>
<tr>
<td>IL-1β (1 ng/ml)</td>
<td>180±27</td>
</tr>
<tr>
<td>IL-1β + 1 μM Res</td>
<td>185±27</td>
</tr>
<tr>
<td>IL-1β + 10 μM Res</td>
<td>203±28</td>
</tr>
<tr>
<td>IL-1β + 100 μM Res</td>
<td>240±27</td>
</tr>
<tr>
<td>IL-1β + 100 μM Q</td>
<td>207±27</td>
</tr>
</tbody>
</table>

Values are means ± SE; n = 6. HDAC, histone deacetylase transferase; HAT, histone acetyltransferase; Res, resveratrol; Q, quercetin.
absence or presence of resveratrol. Cytomix stimulation of these cells at this concentration induces iNOS expression, which can be measured by examining the levels of the NO metabolite nitrite in cell culture medium (16). Resveratrol dose-dependently inhibited cytomix-stimulated release of nitrite (Fig. 8A) and nitrate (NOx; Fig. 8B), with IC50 values of 3.6 ± 2.9 μM (n = 6) and 0.9 ± 0.5 μM (n = 6), respectively. Cytomix stimulated release of GM-CSF (30.9 ± 16.7 and 156.6 ± 39.8 pg/ml in basal and stimulated conditions, respectively) and release of IL-8 (0.66 ± 0.17 and 2.01 ± 0.23 ng/ml in basal and stimulated conditions, respectively) from these cells. Resveratrol also dose dependently inhibited cytomix-stimulated release of GM-CSF and IL-8 from human primary epithelial cells, with IC50 values of 0.44 ± 0.17 and 4.7 ± 3.3 μM, respectively (n = 5; Fig. 8, C and D). Cell viability was unchanged in the presence of increasing concentrations of resveratrol (up to 100 μM) as measured by MTT.

Cytomix exposure of human primary epithelial cells for 4 h led to an induction in iNOS mRNA, and this induction was inhibited by 50 μM resveratrol (Fig. 9, A and B). In contrast, iNOS mRNA was not altered in the presence of the antioxidant N-acetyl cysteine (0.5 mM) or estradiol (50 μM; Fig. 9, A and B). These concentrations were selected because N-acetyl cysteine has been shown to exhibit antioxidant properties on other cell types (14, 30), and if resveratrol was acting as an agonist at the estrogen receptor, the equivalent concentration of estradiol should elicit the same effect. Similarly, the induction of COX-2 mRNA was also reduced by resveratrol, but not by N-acetyl cysteine or estradiol (Fig. 9, A and C). This suggests that the effect of resveratrol cannot be accounted for by its antioxidative or estrogen-like properties. Moreover, resveratrol is not acting as an inhibitor of transcription, because it did not inhibit the basal expression or the twofold induction of tPA with cytomix (Fig. 9, A and D).

**DISCUSSION**

Resveratrol has been ascribed numerous properties that could be beneficial in the treatment of several chronic inflammatory diseases (19). Many studies have been performed using a variety of cell types from different species to investigate the putative mechanisms of action of this molecule; however, no definitive mechanism to explain how resveratrol can exhibit such diverse activities has been identified. Our study is novel, in that it examines the anti-inflammatory mechanism(s) of resveratrol in cells relevant to human disease and explores all the proposed mechanisms in a single study.

Resveratrol exhibited anti-inflammatory activity in all the systems we examined. Furthermore, it appeared to be more effective, although less potent, than glucocorticoids. Resveratrol inhibited inflammatory mediator release from human airway epithelial cells. Release of inflammatory mediators into the cell culture medium was measured as a surrogate for the production of inflammatory mediators into the lung environment, as observed in inflammatory lung diseases including COPD and asthma. Moreover, resveratrol did not act as an estrogen or glucocorticosteroid. In this study, resveratrol inhibited iNOS and COX-2 gene transcription, together with IL-8 and GM-CSF expression in human primary airway epithelial cells. This study confirms the effects of resveratrol on iNOS expression reported in murine macrophage systems (36, 45). However, this contrasts with the effect reported in rat Kupffer cells, where iNOS transcription was unaffected by up to 100 μM resveratrol but translation and, hence, activity were reduced (30). In this study, the expression of iNOS protein was not examined directly; however, iNOS activity was also inhibited in human primary airway epithelial cells. There was also no shift in the balance of nitrate and nitrite formation after inhibition via resveratrol. The inhibition of iNOS expression and activity in primary human airway epithelial cells is significant, because steroids are ineffective in this system (16).

Table 2. Effect of resveratrol, quercetin, and dexamethasone on NF-κB-, TRE-, and CRE-dependent transcription

<table>
<thead>
<tr>
<th>Reporter</th>
<th>Resveratrol</th>
<th>Quercetin</th>
<th>Dexamethasone</th>
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<tbody>
<tr>
<td></td>
<td>EC50 μM</td>
<td>% Inhibition (100 μM)</td>
<td>EC50 μM</td>
</tr>
<tr>
<td>NF-κB</td>
<td>41 ± 10</td>
<td>95 ± 2</td>
<td>21 ± 13</td>
</tr>
<tr>
<td>CRE</td>
<td>46 ± 23</td>
<td>91 ± 3</td>
<td>49 ± 28</td>
</tr>
<tr>
<td>TRE</td>
<td>30 ± 13</td>
<td>89 ± 6</td>
<td>20 ± 12</td>
</tr>
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</table>

Values are means ± SE. CRE, CAMP response element; TRE, TPA response element.
Resveratrol has been shown to inhibit phorbol ester-induced COX-2 transcription in human mammary epithelial cells (42), and this, together with inhibition of NF-κB activation in the monocytic cell line U937 (34), has been purported to be the mechanism for the anti-inflammatory effects of this compound. This study confirms that COX-2 expression and NF-κB activation can also be inhibited in human airway epithelial cells by administration of resveratrol; however, this effect of resveratrol is not specific for NF-κB, inasmuch as resveratrol also inhibited AP-1- and CREB-dependent transcription. Moreover, this is not a general inhibition of inducible gene transcription, because TATA-dependent luciferase activity and expression of tPA in primary cells were unaffected by administration of resveratrol.

There are differences in the efficacy of resveratrol on IL-8 and GM-CSF release from A549 and primary epithelial cells, with inhibition of IL-8 release being less sensitive. There is also a difference in sensitivity between primary epithelial cells and A549 cells, with the cell line being less sensitive to inhibition by resveratrol, despite the use of high concentrations of cytomix required to stimulate the primary cells (16). We previously reported the potency of resveratrol on primary human alveolar macrophages to be less than that observed with A549 cells (12) and nearer to that of primary human airway epithelial cells.

IL-8 and GM-CSF are important in the development of inflammation, because IL-8 is important in the recruitment of...
inflammatory leukocytes, particularly neutrophils, and GM-CSF is a cell survival factor, thus prolonging the resident time of inflammatory cells. The differential inhibitory effects of resveratrol on IL-8 and GM-CSF release further suggest that resveratrol is not simply acting as a general inhibitor of inflammatory mediator release but exhibits some selectivity. The effect of resveratrol and quercetin to inhibit GM-CSF release from A549 cells correlates with their relative efficacies to inhibit NF-κB reporter activity. However, resveratrol and quercetin were more effective at inhibiting AP-1-dependent transcription. The lack of effect of deoxyrhapontin, which is closely related structurally, to inhibit IL-1β-stimulated GM-CSF release from A549 cells indicates that there is a strict, structural requirement for this inhibitory activity. Several studies have demonstrated that the hydroxyl moieties at positions 3 and 5 (Fig. 1) are critical for activity of the resveratrol molecule, and this activity is reduced by substitution with a glucosyl group (2, 36).

It would appear that, despite its structure and reported ability to act as a weak estrogen agonist (20), resveratrol is not acting as an estrogen in this system. In the present study, tamoxifen, an estrogen antagonist, did not antagonize the inhibitory effect of resveratrol in A549 cells. Furthermore, estradiol did not elicit any inhibitory responses in A549 or primary airway epithelial cells. This may relate to the estrogen-independent anti-inflammatory effect of resveratrol reported in MCF-7 breast cancer cells (18). Alternatively, the lack of effect of tamoxifen, an antagonist at estrogen receptor-α and -β, and estradiol in these cells may be due to the reported lack of expression of the estrogen receptor in A549 cells (9, 10); however, type II estrogen binding sites, together with estrogen receptor binding, have been reported in these cells (5). Nevertheless, because estradiol at equivalent concentrations to resveratrol did not attenuate any response in this study, it remains unlikely that the anti-inflammatory effect of resveratrol can be attributed to activation of the estrogen receptor. Similarly, resveratrol is not acting as a glucocorticosteroid, because mifepristone was unable to antagonize resveratrol inhibition of IL-8 release from A549 cells and resveratrol failed to stimulate GRE-dependent luciferase activity. Further evidence suggesting that resveratrol and glucocorticosteroids exert their effects by different mechanisms was demonstrated by the failure of dexamethasone to inhibit the reporter gene assays >50%, whereas resveratrol and quercetin inhibited the responses completely. Resveratrol has also been reported to be more effective than dexamethasone on the adhesion of the monocytic cell line
THP-1 to human umbilical vein endothelial cells via reduced expression of the adhesion molecules ICAM-1 and VCAM-1 on THP-1 cells (2). Recently, resveratrol has been reported to act as an antagonist for the aryl hydrocarbon receptor (6); therefore, the possibility remains that resveratrol could be acting as an agonist/antagonist for other nuclear receptors, the function of which is unknown. More recently, resveratrol has been shown to activate PPAR-α (26); however, the lack of effect of the PPAR-α antagonist to reverse resveratrol-mediated inhibition suggests that it is unlikely that resveratrol is acting via PPAR-α.

A possible mechanism whereby resveratrol and quercetin might be acting as anti-inflammatory agents would be alteration of the HAT-HDAC balance within cells to inhibit inflammatory gene transcription in a manner analogous to the reported effect for glucocorticoids (27). Histone hyperacetylation in Caco-2 human intestinal epithelial cells leads to inhibition of IL-8 release (25). However, resveratrol and quercetin have no effect on the HAT-HDAC balance in A549 cells; therefore, this is an unlikely mechanism for the inhibitory action of their anti-inflammatory effects.

This study demonstrates that plant-derived polyphenolic compounds can act as novel anti-inflammatory agents. Their mechanism of action is not via the estrogen or the glucocorticoid receptor; thus these agents might be beneficial in inflammatory diseases where glucocorticosteroids have proved to be ineffective, such as COPD, steroid-resistant asthma, and arthritis. These compounds may provide candidate molecules for the development of novel anti-inflammatory therapies.

GRANTS
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REFERENCES


