Manganese and iron transport across pulmonary epithelium

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Heilig, Elizabeth A., Khristy J. Thompson, Ramon M. Molina, Alexander R. Ivanov, Joseph D. Brain, and Marianne Wessling-Resnick. Manganese and iron transport across pulmonary epithelium. Am J Physiol Lung Cell Mol Physiol 290: L1247–L1259, 2006.—Pathways mediating pulmonary metal uptake remain unknown. Because absorption of iron and manganese could involve similar mechanisms, transferrin (Tf) and transferrin receptor (TfR) expression in rat lungs was examined. Tf mRNA was detected in bronchial epithelium, type II alveolar cells, macrophages, and bronchus-associated lymphoid tissue (BALT). Tf protein levels in lung and bronchoalveolar lavage fluid did not change in iron deficiency despite increased plasma levels, suggesting that lung Tf concentrations are regulated by local synthesis in a manner independent of body iron status. Iron oxide exposure upregulated Tf mRNA in bronchial and alveolar epithelium, macrophages, and BALT, but protein was not significantly increased. In contrast, TfR mRNA and protein were both upregulated by iron deficiency. To examine potential interactions with lung Tf, rats were intratracheally instilled with 54Mn or 59Fe. Unlike 59Fe, interactions between 54Mn and Tf in lung fluid were not detected. Absorption of intratracheally instilled 54Mn from the lungs to the blood was unimpaired in Belgrade rats homozygous for the functionally defective G185R allele of divalent metal transporter-1, indicating that this transporter is also not involved in pulmonary manganese transport. Pharmacological studies of 54Mn uptake by A549 cells suggest that metal uptake by type II alveolar epithelial cells is associated with activities of both L-type Ca2+/H2O2 channels and TRPM7, a member of the transient receptor potential melastatin subfamily. These results demonstrate that iron and manganese are absorbed by the pulmonary epithelium through different pathways and reveal the potential role for nonselective calcium channels in lung metal clearance.

The iron-binding proteins transferrin (Tf), ferritin, and lactoferrin are also synthesized by pulmonary epithelial cells. These soluble iron-binding factors are found in pulmonary fluid and are thought to contribute to regulation of free iron levels in the lungs and the prevention of oxidative damage (24, 39, 51, 60). The ferroxidase ceruloplasmin is also present in pulmonary fluid and could potentially oxidize Fe2+ to Fe3+, thereby limiting oxidative damage caused by highly reactive ferrous iron and promoting interaction of ferric iron with Tf (39).

Tf not only binds Fe3+, but can also bind Mn3+, albeit with lower affinity (2). Receptor-mediated endocytosis of Tf by transferrin receptor (TfR) expressed on the apical surface of pulmonary epithelial cells therefore could be an important mechanism for clearance of iron, manganese, and other metals from airway and alveolar fluid. DMT1 also plays a key role in TfR-mediated iron uptake as the transporter of iron released from Tf in the endocytic vesicle into the cytoplasm and would be envisioned to have an analogous role in uptake of Mn-Tf via this pathway. As for iron, the speciation of manganese would be an important determinant for transport mediated by DMT1 across the plasma membrane (Mn3+) or for uptake by TfR-mediated endocytosis of Tf-bound metal (Mn4+). Whether
Mn$^{2+}$ would be converted to Mn$^{3+}$ by lung fluid oxidases (e.g., ceruloplasmin) remains unknown.

In addition to the Ti/TIR/DMT1-mediated pathway for metal uptake, there is mounting evidence that nonselective calcium channels transport divalent iron and manganese. Fe$^{2+}$ and Mn$^{2+}$ uptake by cardiomyocytes in vitro (52) and uptake of Fe$^{2+}$ by in situ-perfused heart (33) are both partially blocked by inhibitors of L-type voltage-gated channels (LVGCs) and increased by LVGC activators. Mn$^{2+}$ uptake by liver was found to be sensitive to the LVGC inhibitor nifedipine (12).

Similar analysis of isolated blood-brain-barrier-derived cells, found to be sensitive to the LVGC inhibitor nifedipine (12). This suggests that these metals cross the pulmonary epithelium through different molecular mechanisms (28). Here, we further examine pathways for transport of these metals by pulmonary epithelial cells. Our results demonstrate that after instillation of soluble divalent forms, iron and manganese appear to be cleared from the lungs by distinct mechanisms. Evidence from in vitro studies of Mn$^{2+}$ transport by A549 type II alveolar epithelial cells suggests that nonselective calcium channels may play an important role in uptake of manganese ions by pulmonary epithelial cells.

**MATERIALS AND METHODS**

**Animal care and diets.** Animal protocols for this study were approved by the Harvard Medical Area Animal Care and Use Committee of Harvard University. Male CD/HSD rats (21 days) were purchased from Harlan Sprague Dawley (Indianapolis, IN). Rats were maintained on a 12:12 h light-dark cycle and were given food and water ad libitum. To induce iron deficiency, rats were fed a low-iron diet containing 20–25 ppm Fe for 3 wk (Purina test diet no. 7444; water ad libitum. To induce iron deficiency, rats were fed a low-iron diet containing 20–25 ppm Fe for 3 wk (Purina test diet no. 7444; water ad libitum. To induce iron deficiency, rats were fed a low-iron diet containing 20–25 ppm Fe for 3 wk (Purina test diet no. 7444; water ad libitum. To induce iron deficiency, rats were fed a low-iron diet containing 20–25 ppm Fe for 3 wk (Purina test diet no. 7444; water ad libitum.

Female Dr. M. Garrick (SUNY-Buffalo). Mating pairs of female +/− and male +/− rats were maintained on an iron-supplemented diet containing 500 ppm ferrous iron (TD02385, Harlan Teklad, Madison, WI). Female +/− rats were fed the iron-supplemented diet throughout pregnancy. At postnatal day 6, litters were cross-fostered onto F344 Fischer dams (Harlan Sprague Dawley) fed a standard diet containing 200 ppm iron (Purina diet no. 5053, PharmaServ). On weaning, +/− and +/− rats were fed the iron-supplemented diet (500 ppm). Belgrade (+/−) pups were identified at birth by pale, anemic features and genotype was verified by PCR as described by Fleming et al. (21).

To determine the pharmacokinetics of manganese absorption after intratracheal instillation, +/− and +/− rats were anesthetized with vaporized halothane. 54MnCl$_2$ diluted to 10 μCi/ml in PBS was administered by intratracheal instillation as described above (final delivery volume of 1.5 ml/kg body wt). Blood samples were collected from the tail vein at 5, 15, and 30 min and at 1, 2, and 4 h postinstillation. Rats were killed 1 h postinstillation and brain, heart, liver, lung, bone marrow, muscle, kidney, stomach, small intestine and large intestine were collected for measurement of associated 54Mn. All tissue and blood samples were weighed and radioactivity was measured in a Packard gamma counter (Cobra Quantum, Packard Instrument, IL.), and 54Mn level was calculated as a percentage of the instilled dose.

**In situ hybridization.** After death, the tracheas of control and iron-deficient rats were cannulated with a blunt 18-gauge needle and syringe containing optimum cutting temperature (OCT) compound (Sakura Finetek USA, Torrance, CA) prewarmed to 37°C. Lungs were filled with OCT, cut laterally into sections, mounted in OCT, snap-frozen in 2-methylbutane chilled on dry ice, and stored at −80°C. Ten-nanometer-thick sections were cut on a cryotome and stored at −20°C until processed. Digoxigenin-labeled sense and antisense cRNA probes were transcribed from a full-length rat Ti cDNA (generously supplied by Dr. S. Hisayasu, Tokyo, Japan) or a 594-bp rat TIR EST (Resgen/Invitrogen, Carlsbad, CA), flanked by T7 and T3 promoter sites. Transcripts were shortened to an average length of 200–400 bp by alkali hydrolysis. Sections were incubated with sense or antisense probe (~200 ng/ml) in hybridization buffer (50% formamide, 5% SSC, 2% blocking reagent, 0.02% SDS, 0.1% N-laurylsarcosine). Hybridized probes were detected using anti-digoxigenin-alkaline phosphatase-coupled Fab fragments and bromo-4-chloro-3-indoxyl-phosphate and nitro blue tetrazolium as a substrate. Sections were incubated in substrate solution for 42 h, rinsed in TE buffer (10 mM Tris and 1 mM EDTA, pH 8.0), and mounted in 50% PBS-glycerol.

**Detection of proteins by Western blotting.** Snap-frozen lung tissue from control, Fe-deficient, or Fe oxide-exposed rats was homogenized on ice in 20 mM HEPES, pH 7.4, 100 mM KCl, 85 mM sucrose, 20 μM EGTA, 1% Triton X-100, with Complete Mini protease inhibitor cocktail (Roche, Indianapolis, IN) added. Homogenates were incubated at 4°C with rotation for 45 min and then clarified by centrifugation for 10 min at 12,000 g. Protein concentration in supernatants was measured by Bradford assay (6). Samples of 120 μg lung protein or 20 μg BAL protein were electrophoresed on 10 or 7% SDS-polyacrylamide gels, respectively, and transferred to nitrocellulose for...
immunoblotting. Membranes were blocked with 5% nonfat dry milk in TBST (50 mM Tris, 150 mM NaCl, 0.1% Tween 20, pH 7.4) for 1 h at room temperature, then incubated with mouse anti-human TIR (1:1,000; Zymed, San Francisco, CA), rabbit anti-human TF (1:2,000, Rockland), sheep anti-human ferritin (1:1,000, The Binding Site, Birmingham, UK) or mouse anti-α-tubulin (1:10,000, Sigma, St. Louis, MO) overnight at 4°C. Secondary horseradish peroxidase-labeled goat anti-mouse IgG (1:40,000, Pierce, Rockford, IL) or donkey anti-rabbit IgG (1:40,000, Pierce) was used to detect immunoreactivity by enhanced chemiluminescence using SuperSignal West Pico Reagent (Pierce). Intensity of bands was quantified with Bio-Rad QuantityOne software.

FPLC fractionation of BAL fluid. After intratracheal instillation of 54Mn or 55Fe as described above, BAL supernatant was centrifuged at 12,000 g for 15 min to remove insoluble material and debris. Two milliliters were then dialyzed (Spectra/Por, MWCO 12,000–14,000; Spectrum Laboratories, Rancho Dominguez, CA) against FPLC buffer A (25 mM Tris, pH 8.5) for 4–6 h at 4°C to lower salt concentration. Samples were loaded on a MonoQ anion-exchange column and proteins were eluted with a twostep linear salt gradient (40 ml of 0–0.5 M NaCl, followed by 20 ml 0.5–1.0 M NaCl). Radioactivity in 1 ml fractions from FPLC, along with cpm in lungs, BAL cell pellet, and BAL (pre- and postdialysis) was measured by gamma counting.

For FPLC fractionation of 54Mn-Tf, a 100-μl solution containing 12.5 μM human apo-Tf (Sigma), 125 μM MnCl2, and 125 mM 54MnCl2 (0.3 μCi) in 15 mM NaHCO3 (pH 8.0) was incubated at 37°C for 5 h. The binding of manganese to Tf was confirmed by measuring the absorbance ratio at 300 nm/280 nm (16), and 54Mn-Tf was subsequently dialyzed (molecular weight cutoff 12,000–14,000) against FPLC buffer A for 12–16 h at 4°C. 54Mn-Tf was fractionated by FPLC on a MonoQ column exactly as described above.

Protein fractions were precipitated overnight at 4°C (25% trichloroacetic acid, 0.02% casamino acids), washed twice with ice-cold acetic acid, and resuspended by boiling for 10 min in 1× Laemmli buffer containing 5% β-mercaptoethanol. Fractions were electrophoresed on 5–20% SDS-polyacrylamide gels and proteins were visualized by either Coomassie blue or silver staining. Bands of interest were excised and digested overnight at 37°C in 50 mM NH4HCO3 containing 10–20 μg/ml trypsin. Tryptic peptides were extracted with one change of 20 mM NH4HCO3, two changes of 5% formic acid in 50% acetonitrile, and one change of 5% formic acid in 50% isopropanol at room temperature. For identification of proteins by nanoliquid chromatography (LC) electrospray ionization tandem mass spectrometry, digests were injected onto a C18 solid-phase extraction column for desalting, eluted onto and separated on a nano-LC column with a linear gradient of acetonitrile in 0.1% formic acid, and the eluent was introduced into a LCQ Deca XP Plus mass spectrometer by nanoelectrospray. A full MS scan between 400 and 1,800 m/z was performed, followed by five MS/MS scans for the five most intense ions from the MS scan acquired in data-dependent MS/MS scanning experiments. Data files were converted to .dta files using Bioworks 3.2 software (ThermoElectron) and submitted to the database search against a composite database (forward and reversed [40]) protein sequence database derived from NCBI rat database on a Sorecer search station (Sage-N, Thermo Electron, Philadelphia, PA) using SEQUEST algorithm. SEQUEST output (.out files) was analyzed using DTASelect software (provided by Dr. J. Yates, Scripps Research Institute).

54Mn and 55Fe uptake assays. A549 cells (ATCC no. CCL-185; kindly provided by Dr. D. Tschumperlin, Harvard School of Public Health) were maintained in DMEM/F12 (1:1) media (Mediatech, Herndon, VA) supplemented with 10% fetal bovine serum (Gemini Bio Products, woodland, CA) and seeded at a density of 3.6 × 105 cells per well in 6-well plates, or 1.6 × 106 cells per well in 12-well plates, 24 h before uptake assays. Cells were subconfluent (70–80%) at time of assays. Cells were washed 3× with HeBS (20 mM HEPES, 137 mM NaCl, 0.5 mM Na2HPO4, 0.5 mM KCl, pH 7.4) and then incubated with 1–2 μCi 54MnCl2 (Perkin Elmer/New England Nuclear Corp.) in HeBS with drugs or vehicle for 4 min at 37°C unless otherwise indicated. After being washed 3× with HeBS, cells were incubated with 100 mM MnCl2 in HeBS for 1 h at 4°C to eliminate nonspecific binding of 54Mn. Cells were then washed 3× in HeBS and lysed in 600 μl of solubilization buffer (0.02 M NaOH, 0.1% Triton X-100). Simultaneous control assays were performed at 4°C for all treatments. For assays with 55Fe, a stock solution of 55Fe-nitrotoluene-acetic acid (1:4) was diluted with a 1:50 molar ratio of freshly made ascorbic acid immediately before each assay, then diluted further in HeBS uptake buffer at indicated pH. Radioactivity of lysates was measured in a gamma (54Mn) or beta (55Fe) counter, and protein concentration was determined by Bradford assay. Specific uptake was taken as the difference between uptake measured at 37 and 4°C and was calculated as picomole Mn per microgram of protein. MgCl2, NiCl2, and CaCl2 stocks were 0.1–1.0 M in H2O. FeCl3 was dissolved in water and mixed with a 1:50 molar excess of ascorbic acid immediately before use. Drug stocks were prepared as follows: 10 mM in H2O; amiloride, 10 mM in DMSO; thapsigargin, 3 mM in DMSO; nifedipine, 10 mM in ethanol; BAY K 8644, 0.1 mM in ethanol; 2-aminoethoxydiphenyl borate (2-APB), 10 mM in DMSO; and oleylacteylglycerol (OAG), 20 mM in DMSO.

RT-PCR. RNA was isolated from subconfluent A549 cells with RNA- Bee (Tel-Test, Friendswood, TX) following the manufacturer’s instructions. RNA was treated with DNase I (Promega, Madison, WI) to remove genomic DNA, extracted with phenol-chlororm-isooamyl alcohol (25:24:1) mixture, precipitated, resuspended in diethyl pyrocarbonate-treated water, and quantified by UV spectrophotometry. cDNA was synthesized from 2 μg RNA in reactions containing 1× Moloney murine leukemia virus-reverse transcriptase (MMLV-RT) buffer, 0.5 mM dNTP mix, 25 μg/ml oligodT15 primer, 5 mM MgCl2, 10 mM dithiothreitol, 1 U of RNasin, and 200 U of MMLV-RT enzyme (Promega). MMLV-RT enzyme was omitted for RT-control reactions. Amplification reactions were carried out with 1 μl of cDNA in 1× PCR buffer (10 mM Tris–HCl, pH 9.0, 50 mM KCl, 2.0 mM MgCl2; Promega). Sequences of primers for TRPC1–6 were previously published elsewhere (53). Sequences of other primers used are as follows: TRPC7 forward, 5'-CTGCCAGCACAGGATGAG-3'; TRPC7 reverse, 5'-TCATCAAGTGAACTAGCAGGATG-3'; TRPM7 forward, 5'-TCCAAGCACGTATTTAAAACGGTCATAA-3'; TRPM7 reverse, 5'-GTCCTAGCAGCTAGCTGTCG-3'; LVGC (α-subunit) forward, 5'-AGAATCTGGGGCAGAGTATGAC-3' and LVGC reverse, 5'-CAGGAAGCATTGGCGTAAGAC-3'.

Fifteen microliters of each reaction were separated on a 1.2% agarose gel containing ethidium bromide, and band intensity was quantified with QuantityOne software (Bio-Rad, Hercules, CA).

Statistical analysis. All values are means ± SE. Comparisons of protein expression measured by Western blotting and A549 cell metal uptake assays were evaluated by unpaired Student’s t-test. Characteristics of the pharmacokinetics of manganese transport and tissue distribution of 54Mn were evaluated by multivariate ANOVA (MANOVA) using the general linear model procedure (SAS statistical analysis software; SAS Institute, Cary, NC). Statistical significance was based on an α level of 0.05.

RESULTS

Expression of transferrin in lungs. Expression and iron-responsive regulation of Tf and TIR were examined by in situ hybridization and Western blot analysis of lung tissue from control, iron-deficient, and iron oxide-exposed rats. In control animals, Tf mRNA was detected in type II alveolar epithelial cells, alveolar macrophages, bronchial epithelium, and in localized regions of bronchus-associated lymphoid tissue (BALT; Fig. 1, A and D). Tf mRNA expression in iron-deficient rats exhibited the same localization pattern and stain-
ing intensity as in controls (Fig. 1, B and E). In iron oxide-exposed lung, staining for Tf mRNA was greater in BALT and bronchial epithelium than in control lung (Fig. 1F) and appeared more intense in alveolar epithelial cells and macrophages in regions where iron particles were deposited (Fig. 1C).

Western blot analysis indicated that Tf protein levels were not significantly altered in lung tissue from either iron-deficient or iron oxide-exposed rats. To examine whether Tf levels in pulmonary fluid change in iron deficiency, protein levels in BAL fluid from control and iron-deficient rats were quantified by Western blotting. While plasma levels were significantly elevated in iron-deficient rats as expected, the level of Tf in BAL was not significantly different from controls (Figs. 2 and 3). Because circulating plasma Tf levels are significantly elevated in dietary iron deficiency (5), the apparent lack of change in BAL Tf levels in iron-deficient animals suggests that the concentration of this protein in pulmonary fluid reflects synthesis by the lungs (60) in a manner that is independent of body iron status.

Expression of transferrin receptor in lungs. TfR mRNA was detected only in BALT and bronchial epithelium of control rats (Fig. 4, A and D). In iron-deficient rats, similar staining intensity was observed in BALT and bronchial epithelium (Fig. 4E), while upregulation of TfR message was observed in type II alveolar epithelial cells (Fig. 4B). In contrast, staining in iron oxide-exposed rats was significantly stronger in BALT and bronchial epithelium (Fig. 4F) and was also increased in alveolar epithelium and in macrophages in areas of iron oxide particle deposition (Fig. 4C).

Western blot analysis to examine iron-responsive changes in lung TfR indicated that protein levels were significantly increased in iron-deficient rat lung (Fig. 2). In contrast to mRNA levels, however, the level of TfR protein did not change in iron...
oxide-exposed lung. Ferritin levels were significantly higher in iron oxide-exposed lung, confirming increased tissue iron levels.

Interaction of $^{54}$Mn and $^{59}$Fe with proteins in BAL fluid. To determine whether soluble manganese or iron introduced into the lungs interacts with Tf in pulmonary fluid, rats were intratracheally instilled with $^{54}$Mn or $^{59}$Fe and lavage fluid was collected 10 min postinstillation. Although the levels associated with lung tissue were comparable for $^{54}$Mn and $^{59}$Fe (~55% instilled dose), significant differences were observed in the distribution of the radioisotopes in BAL fluid (Table 1). Only ~14% of instilled $^{54}$Mn remained in BAL fluid after 10 min of which 1% was found to be associated with protein (as determined by dialysis). In contrast, 27% of instilled $^{59}$Fe was found in BAL, of which 22% was bound to protein. To examine the association of the radioisotopes with Tf in lung fluid, in vivo-radiolabeled BAL samples were fractionated by anion-exchange chromatography using fast-performance liquid chromatography. The two major chromatographic peaks were identified as Tf (fraction 17) and albumin (fractions 26–27) by Western blotting (data not shown). After dialysis of BAL fluid, $^{59}$Fe was found predominantly in fraction 17 containing the peak for Tf (Fig. 5A). In contrast, $^{54}$Mn was detected only in fraction 15 (Fig. 5B). Fractionation of BAL fluid collected 1 h after instillation of $^{54}$Mn demonstrated that only trace amounts of $^{54}$Mn are associated with protein in pulmonary fluid after

![Fig. 2. Transferrin receptor (TfR), Tf, and ferritin protein levels in control, Fe-deficient, and Fe oxide-exposed rat lung. A: Western blot analysis was performed as described under MATERIALS AND METHODS. Lanes contain samples from individual animals. Separate experiments were conducted for Fe-deficient and Fe oxide-exposed groups, with age-matched controls. Note that ferritin levels were below limit of detection in control and Fe-deficient lung. B: Tf and TfR protein expression levels in A were quantified (QuantityOne imaging software, Bio-Rad). Values are expressed as % control and normalized to α-tubulin expression level. *P < 0.0005 (n = 4 to 5; means ± SE).](http://ajplung.physiology.org/)

![Fig. 3. Tf levels increase in plasma but not bronchoalveolar lavage (BAL) of iron-deficient rats. A: Western blot analysis was performed to determine Tf levels in BAL and plasma of control and Fe-deficient rats. B: band intensity detected by chemiluminescence was quantified using QuantityOne imaging software (Bio-Rad). *P < 0.05 (n = 5 to 6; means ± SE).](http://ajplung.physiology.org/)
longer in vivo incubation periods (Fig. 5C). To demonstrate that $^{54}$Mn bound to Tf would remain associated during fractionation, apoTf was loaded with $^{54}$Mn in vitro and subjected to dialysis and anion exchange FPLC procedures. In vitro-radiolabeled $^{54}$Mn-Tf was only detected in fraction 17, confirming that if present in BAL fluid, this species would be detected under our experimental conditions (Fig. 5D).

To identify the protein(s) that might bind $^{54}$Mn in fraction 15, samples were pooled and concentrated. Six bands were detected by silver staining and identified by mass spectrometry (Fig. 6B). Of the proteins identified, only transketolase peaked in fraction 15. Transketolase is also the only protein identified in this fraction, other than Tf, that has been shown to bind manganese (30). Transketolase is a cytoplasmic enzyme that catalyzes transfer of dihydroxyethyl groups from ketose donors to aldose acceptors in the pentose phosphate pathway and is not predicted to function in ion transport (49).

Table 1. Distribution of $^{54}$Mn and $^{59}$Fe after intratracheal instillation

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<thead>
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<th>$^{54}$Mn, % Instilled Dose</th>
<th>$^{59}$Fe, % Instilled Dose</th>
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<tbody>
<tr>
<td>BAL (bound to protein)*</td>
<td>1.15±0.29</td>
<td>22.24±5.15</td>
</tr>
<tr>
<td>BAL (unbound)</td>
<td>13.40±3.20</td>
<td>5.29±1.62</td>
</tr>
<tr>
<td>BAL cell pellet</td>
<td>0.70±0.13</td>
<td>0.75±0.14</td>
</tr>
<tr>
<td>Lung tissue</td>
<td>55.01±3.38</td>
<td>54.08±8.17</td>
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<tr>
<td>Total in lungs</td>
<td>70.25±0.87</td>
<td>82.36±1.46</td>
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Values are means ± SE, n = 3. *Refers to radioactivity in bronchoalveolar lavage (BAL) after dialysis with molecular weight cutoff ≥ 14,000.

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Pharmacokinetics of $^{54}$Mn absorption by Belgrade rats after intratracheal instillation. Because an association between instilled $^{54}$Mn and Tf could not be detected in vivo, our findings suggested the possibility that soluble divalent manganese might be taken up directly by the lungs after instillation. One transporter that might be involved in pulmonary Mn$^{2+}$ uptake is DMT1 (26, 56). Homozygous Belgrade (b/b) rats with defective DMT1 function exhibit reduced clearance of iron and vanadium from the lungs after instillation with ROFA com-
pared with heterozygous (+/b) controls (25). Therefore, we examined the pharmacokinetics of $^{54}$Mn transport from the lungs to the blood after intratracheal instillation of homozygote (b/b) Belgrade rats and heterozygote (+/b) siblings. Figure 7 shows isotope levels determined in blood samples drawn from 5 min to 4 h postinstillation for both experimental groups. As previously observed (28), lung manganese absorption was quite rapid, with an initial drop in blood levels within 30 min postinstillation and relatively stable blood levels thereafter. The pharmacokinetics did not differ between b/b and b/+ rats. At 4 h, circulating levels of $^{54}$Mn in homozygous Belgrade and control heterozygous rats were similar (0.14 and 0.09%, respectively). The tissue distribution of $^{54}$Mn determined 4 h postinstillation is shown in Table 2. The amounts of $^{54}$Mn remaining in the lungs of b/b and b/+ rats at this time point were also similar (60.73 and 64.09%, respectively), further indicating that pulmonary absorption of manganese was not impaired in Belgrade rats. The only differences noted in tissue distribution of $^{54}$Mn after intratracheal instillation were that levels were significantly higher in the spleen and large intestine and lower in the kidneys of b/b rats compared with their littermate b/+ controls (Table 2).

Characterization of $^{54}$Mn uptake by A549 cells. To examine pathways for direct uptake of divalent manganese by the lungs, transport of $^{54}$Mn$^{2+}$ was characterized using A549 cells, a type II alveolar epithelial cell line. $^{54}$Mn$^{2+}$ uptake was time and temperature dependent (Fig. 8A). Uptake was also saturable with increasing Mn concentration (Fig. 8B). Uptake of $^{55}$Fe$^{2+}$ by A549 cells was increased at lower pH; however, $^{54}$Mn$^{2+}$ uptake at pH 6.75 was comparable to uptake observed at pH 7.4 (Fig. 9). These results suggest that DMT1 may be involved in ferrous iron uptake by alveolar epithelial cells as this membrane protein functions as a proton symporter, exhibiting increased Fe$^{2+}$ and Mn$^{2+}$ transport activity at pH 6.75 (22, 57). While the lack of a pH effect on A549 cell $^{54}$Mn uptake does not rigorously rule out a possible role for DMT1 in transport of this metal in the lungs, it is in general agreement with the observations that uptake of intratracheally instilled $^{54}$Mn is unimpaired in the Belgrade rat model.

Effect of calcium channel antagonists and agonists on $^{54}$Mn uptake. We next examined whether Ca$^{2+}$, Fe$^{2+}$, or Ni$^{2+}$ would compete with Mn for uptake by A549 cells (Fig. 10A). The observation that the presence of 500 μM Ca$^{2+}$ inhibited uptake of $^{54}$Mn suggested one or more calcium channels could be involved in the influx pathway. Coincubation with a 10- or 100-fold excess of Ni$^{2+}$, a broad-spectrum inhibitor of both voltage-gated and store-operated calcium channels, inhibited $^{54}$Mn uptake by 69 and 86%, respectively. Likewise, dose-dependent inhibition by Fe$^{2+}$ was observed with a 100-fold excess resulting in ~93% inhibition relative to uptake in control wells.

Pharmacological characteristics of various drugs with specificity for different Ca$^{2+}$ channels suggested both voltage-gated channels (VGCS) and receptor-operated channels (ROCIs), but not SOCs, are involved in manganese uptake by A549 cells (Fig. 10B). Treatment of cells with amiloride, an inhibitor with higher specificity for T-type VGCS, resulted in a relatively modest inhibition of transport (~10%). In contrast, the inhibitor verapamil, which acts on L-type VGCS, substantially reduced uptake and acted in a dose-dependent manner with ~65% inhibition at 500 μM. The dihydropyridine (DHP)
inhibitor nifedipine and the DHP activator \( S(-)\)-BAY K 8644, which bind to a specific site on the \( \alpha_1 \)-subunit of L-type VGCs, had no effect on \(^{54}\text{Mn}\) uptake. Thapsigargin, which causes release of \( \text{Ca}^{2+} \) from intracellular stores thereby activating SOCs, also had no effect. 2-APB, an inhibitor of the ROCs TRPC3, 6, and 7, reduced transport; however, the diacylglycerol analog OAG, which is reported to stimulate \( \text{Ca}^{2+} \) uptake by these members of the TRPC family (19, 29, 61), had no effect on \(^{54}\text{Mn}\) uptake. Coincubation with 4 \( \text{mM MgCl}_2 \), which inhibits transport of divalent cations by TRPM7 (35), effectively decreased \(^{54}\text{Mn}\) uptake by 75%.

Identification of TRP channels expressed by A549 cells. Electrophysiological studies in which \( \text{Mn}^{2+} \) was substituted for \( \text{Ca}^{2+} \) have demonstrated that several members of the TRPC (transient receptor potential canonical) subfamily are perme-
TRPM7, a member of the TRPM (melastatin) subfamily, is also permeable to several divalent metal ions, including Mn$^{2+}$. To determine which TRP channels are expressed by A549 cells, and to verify expression of LVGCs, RT-PCR was performed (Fig. 11). Expression of TRPC1, 3, 6, and 7, TRPM7, and the α-subunit of LVGC was detected. Interestingly, primers for TRPC7 amplified two products, one of the expected size (428 bp), and a smaller amplicon of 200 bp. Because the 5′ and 3′ primers are located in different exons (exons 1 and 2, respectively), the 200-bp amplicon may be the result of alternative splicing of TRPC7 mRNA in A549 cells.

**DISCUSSION**

Manganism can result from inhalation of manganese-laden dust, making the lungs an important route of exposure to this potentially toxic metal. In this study, we investigated the molecular mechanisms of manganese uptake by pulmonary epithelial cells. Tf is reported to bind Mn in plasma (18) and is highly abundant in pulmonary fluid. In iron deficiency, circulating plasma Tf levels increase dramatically, and TfR expression is posttranscriptionally upregulated by iron regulatory protein (IRP) activation (11). Conversely, iron overload results in downregulation of TfR, coupled with increased expression of the cytosolic iron storage protein ferritin. Because Tf and TfR expression levels in the lungs could potentially affect clearance of inhaled metals, we examined expression of mRNA and protein in rats with iron deficiency and in rats exposed to iron oxide particles. In agreement with a previous report (60), our study shows that Tf mRNA is widely expressed by pulmonary cells, including bronchial epithelium, type II alveolar epithelial cells, macrophages, and focal regions of BALT (Fig. 1). Expression of Tf mRNA or protein in lung and BAL fluid did not change in iron deficiency, demonstrating that synthesis of Tf by pulmonary cells is regulated in response to the local environment of the lungs, rather than by systemic responses to iron status. Previous studies from our laboratory showed that iron deficiency does not enhance absorption of $^{59}$Fe from the lungs and are consistent with these data (28). Iron oxide particles induced upregulation of Tf mRNA in bronchial and alveolar epithelium, macrophages, and BALT but protein levels did not change, although a nonsignificant trend toward increased Tf protein was detected in whole lung lysate.

TfR mRNA was detected primarily in BALT, with weak staining in bronchial epithelium. TfR mRNA expression was...
enhanced in type II alveolar epithelial cells by iron deficiency, and TfR protein levels in whole lung were significantly increased. Iron oxide exposure also resulted in induction of TfR mRNA in lung; however, TfR protein levels did not change. Because ferritin was upregulated in iron oxide-exposed lung, it is unlikely that IRP activity accounts for posttranscriptional regulation of TfR mRNA. These findings suggest that while TfR gene expression may be induced at the transcriptional level by inflammatory stimuli (54), protein expression of lung TfR does not change when iron concentration is very high as a defense against cytotoxicity.

To test whether the Tf/TfR pathway might be involved in uptake of manganese, 54Mn2+ was administered to rats by intratracheal instillation. At 10 min postinstillation, ~85% of the dose of 54Mn had been absorbed from pulmonary fluid and only a small fraction (~1%) of the remaining 54Mn was associated with BAL fluid protein. In contrast, 27% of the dose of intratracheally instilled 59Fe2+ remained in BAL fluid, the majority of which was associated with Tf. These data suggest that while soluble 59Fe2+ is oxidized to Fe3+ after instillation into the lungs and rapidly binds to Tf, 54Mn2+ is directly taken up by pulmonary epithelial cells via Tf-independent pathways. This idea is confirmed by the fact that 1 h postinstillation, very little of the instilled 54Mn remains bound to BAL fluid protein. It is likely that association of instilled soluble iron with Tf is oxidized to Fe3+ after instillation into the lungs and rapidly binds to Tf, 54Mn2+ is directly taken up by pulmonary epithelial cells via Tf-independent pathways. This idea is confirmed by the fact that 1 h postinstillation, very little of the instilled 54Mn remains bound to BAL fluid protein. It is likely that association of instilled soluble iron with Tf

Fig. 10. Pharmacological profile of 54Mn uptake by A549 cells. A: cells were incubated with 0.68 μM 54Mn2+ in the presence or absence of other divalent cations at the concentrations indicated. B: transport assays with 0.34 μM 54Mn2+ in the presence and absence of inhibitors/activators of L-type calcium channels, TRPC channels, and TRPM7. Cells were incubated with 54Mn ± treatment or vehicle for 4 min. *P < 0.05 (n = 3–4; means ± SE).

Although DMT1 is known to play a role in the endocytic Tf/TfR pathway, the function of this divalent metal transporter at the plasma membrane could also provide a potential pathway for the direct uptake of Mn2+. However, two separate lines of evidence argue against this possibility. In vivo experiments with Belgrade rats demonstrate that the impaired DMT1 activity of homozygous animals does not modify the pharmacokinetics of pulmonary 54Mn absorption from the lungs to the blood. Moreover, in vitro studies of 54Mn2+ uptake by A549 cells show that transport is not enhanced by low pH, indicating that DMT1 cannot be the predominant Mn transport activity. These results prompted us to consider alternative pathways for pulmonary manganese uptake. Pulmonary epithelial and endothelial cells express several calcium channels that are essential in maintenance of fluid balance in the lungs (34). Several recent publications have demonstrated that uptake of manganese into heart, liver, and brain is sensitive to modulators of LGVGCs (12, 17, 52). Furthermore, patch-clamp studies have shown that several members of the TRPC subfamily of non-selective calcium channels are permeable to Mn2+ (9, 29). TRPC proteins are store-operated and/or receptor-operated calcium channels that can be activated by intracellular store depletion and/or by stimulation of G protein- or tyrosine kinase-coupled receptors (36). Formation of heteromultimeric complexes within subfamilies (e.g., TRPC4/5, TRPC3/6, TRPC1/3/7) produces channels with different characteristics and contributes to functional variability in vivo (48). Whether TRPC channels are permeable to Mn2+ in vivo is not clear. A
widely expressed member of the TRP melastatin-related (TRPM) subfamily, TRPM7, is also implicated in Mn\(^{2+}\) transport. TRPM7 is important in cellular magnesium homeostasis and is involved in human disorders of magnesium metabolism (10). TRPM7 transports several divalent metal ions under physiological conditions, including Mn\(^{2+}\) (35, 37).

Analysis of gene expression of TRPC family members by RT-PCR and Northern blotting has shown that TRPC1, 3, 4, 5, 6, and 7, and TRPM7 are expressed in lung (43, 45). TRPC4 and 6 have been found in differentiated human bronchial epithelial cells, whereas TRPC3 and TRPM7 are expressed by bronchial epithelium-derived BEAS-2B cells (10, 13). Expression of TRPC1 and TRPC6 in A549 cells was previously reported (59). We have verified by RT-PCR that A549 cells express TRPC1, TRPC6, and LVGC and have additionally determined that TRPC3, TRPC7, and TRPM7 are also expressed in this cell line.

Pharmacological features of manganese transport in A549 cells were defined in our study by the effects of several calcium channel inhibitors and activators on \(^{54}\)Mn\(^{2+}\) uptake. Treatment with verapamil, a phenoxyalkylamine antagonist that binds to the \(\alpha_1\)-subunit of LVGC, caused a significant dose-responsive reduction in \(^{54}\)Mn uptake. Treatment with the DHP antagonist nifedipine or the activator BAY K 8644, both of which bind to a specific site on the \(\alpha_1\)-subunit distinct from the binding site of verapamil, did not have an effect. Because the efficacy of DHP drugs appears to be voltage dependent, their lack of effect in A549 cells could be due to the fact that these cells are at resting potential (4, 47). Alternatively, verapamil could potentially inhibit Mn influx through TRPC3 (63) or by another uncharacterized mechanism.

To determine whether manganese influx might also occur through SOCs, including TRPC1 and TRPC7 (31), A549 cells were treated with thapsigargin, an inhibitor of sarcoplasmic/endoplasmic reticulum Ca\(^{2+}\) ATPase that produces a rapid increase in cytosolic [Ca], triggering opening of SOCs on the plasma membrane. Treatment with thapsigargin has previously been shown to increase Ca\(^{2+}\) influx and trigger Mn\(^{2+}\)-mediated quenching of fura-2 fluorescence in A549 cells (59). We did not observe any effect of thapsigargin on \(^{54}\)Mn uptake, but we note that the concentration of Mn\(^{2+}\) studied by Xue et al. (59) was 1.2 mM, ~2,000-fold greater than the concentrations used in our assays.

2-APB is a noncompetitive inhibitor of the \(\mathrm{I}_3\) receptor which blocks activation of ROCs in response to G protein-coupled receptor (GPCR)-mediated phospholipase C (PLC) activation (32). 2-APB can also act directly on TRPC3, TRPC5, TRPC6, and TRPC7, possibly by binding in the pore region of these channels (31, 58). Inhibition of \(^{54}\)Mn uptake by 2-APB suggests involvement of one or more of the 2-APB-sensitive TRPC proteins that behave as ROCs. However, we found that OAG, a membrane-permeable diacylglycerol (DAG) analog that activates TRPC3, 6, and 7, had no effect on \(^{54}\)Mn uptake. The subunit composition of TRPC heteromeric channels in vivo modulates sensitivity to DAG (50) and could explain this apparently contradictory finding. A second explanation might be that TRPM7 is responsible for manganese uptake. One report has indicated that calcium influx through TRPM7 is also sensitive to 2-APB treatment (27). Unlike the TRPC channels, TRPM7 is inhibited by PLC-mediated depletion of PIP\(_2\) following GPCR stimulation and is not activated by OAG (1, 46). Depletion of intracellular Mg\(^{2+}\) and Mg-ATP activates TRPM7, and high concentrations of either form of magnesium inhibit channel activity (38). In our assays, addition of 4 mM MgCl\(_2\) resulted in a 75% inhibition of \(^{54}\)Mn\(^{2+}\) uptake by A549 cells, supporting a key role for this channel. The preference of TRPM7 for manganese and other divalent metals over calcium make this channel a plausible candidate for manganese transport in vivo (35).

In conclusion, transition metals in ambient particulate matter from diesel exhaust, ROFA, and coal dust are thought to be the component(s) responsible for increased airway hyperresponsiveness, inflammation, and oxidative stress from these exposures (23, 42). We have characterized changes in Tf and TiR expression in the lungs in response to iron status that may reflect how the lungs handle metal burden to minimize oxidative damage. Of equal importance are the mechanisms by which potentially toxic inhaled metals enter the blood stream. While a role for lung fluid Tf in iron absorption should be tested more rigorously, our results suggest that ferrous iron can be readily oxidized to bind Tf and that elements of the Tf/TiR/DMT1 pathway may mediate its absorption from the lungs. In contrast, our instillation studies indicate that soluble Mn\(^{2+}\) does not interact with Tf in BAL fluid, suggesting alternative pathways for uptake are responsible for its transport from the lungs. One caveat is that the potential role for the Tf/TiR/DMT1 pathway in the uptake of trivalent Mn cannot be rigorously ruled out as it is possible that on dissolution, metal from particulate matter could be oxidized and cleared by this pathway. Our findings do support a potential role for TRPM7, LVGCs, and possibly other TRP channels in the process of lung Mn\(^{2+}\) absorption. It will be of great interest to determine whether such channels function in transport of trace metals in the lungs in vivo and whether exposure to components of particulate matter and/or resulting inflammation may affect channel activity.

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DISCLOSURES

This work has not been formally reviewed by either the NIEHS or EPA. The views expressed in this document are solely those of the authors and do not necessarily reflect those of either the NIEHS or the EPA. Neither NIEHS nor the EPA endorses any products or commercial services mentioned in this publication.

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