Angiotensin II type 2 receptor ligand PD123319 attenuates hyperoxia-induced lung and heart injury at a low dose in newborn rats

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1Department of Pediatrics, Division of Neonatology, Leiden University Medical Center, Leiden, The Netherlands; 2Division of Vascular Disease and Pharmacology, Department of Internal Medicine, Erasmus Medical Center, Rotterdam, The Netherlands; 3Department of Pharmacology, Utrecht Institute for Pharmaceutical Sciences, Utrecht University, Utrecht, The Netherlands; and 4Department of Pediatrics, Los Angeles Biomedical Research Institute at Harbor-UCLA Medical Center, Torrance, California

Submitted 27 November 2013; accepted in final form 13 June 2014

Wagenaar GT, Sengers RM, Laghmani EH, Chen X, Lindeboom MP, Roks AJ, Folkerts G, Walther FJ. Angiotensin II type 2 receptor ligand PD123319 attenuates hyperoxia-induced lung and heart injury at a low dose in newborn rats. Am J Physiol Lung Cell Mol Physiol 307: L261–L272, 2014. First published June 20, 2014; doi:10.1152/ajplung.00345.2013.—Intervening in angiotensin (Ang)-II type 2 receptor (AT2) signaling may have therapeutic potential for bronchopulmonary dysplasia (BPD) by attenuating lung inflammation and preventing arterial hypertension (PAH)-induced right ventricular hypertrophy (RVH). We first investigated the role of AT2 inhibition with PD123319 (0.5 and 2 mg·kg–1·day–1) on the beneficial effect of AT2 agonist LP2–3 (5 μg/kg twice a day) on RVH in newborn rats with hyperoxia-induced BPD. Next we determined the cardiopulmonary effects of PD123319 (0.1 mg·kg–1·day–1) in two models: early treatment during continuous exposure to hyperoxia for 10 days and late treatment starting on day 6 in rat pups exposed postnatally to hyperoxia for 9 days, followed by a 9-day recovery period in room air. Parameters investigated included lung and heart histopathology, fibrin deposition, vascular leakage, and differential mRNA expression. Ten days of coadministration of LP2–3 and PD123319 abolished the beneficial effects of LP2–3 on RVH in experimental BPD. In the early treatment model PD123319 attenuated cardiopulmonary injury by reducing alveolar septal thickness, pulmonary influx of inflammatory cells, including macrophages and neutrophils, medial wall thickness of small arterioles, and extravascular collagen III deposition, and by preventing RVH. In the late treatment model PD123319 diminished PAH and RVH, demonstrating that PAH is reversible in the neonatal period. At high concentrations PD123319 blocks the beneficial effects of the AT2-agonist LP2–3 on RVH. At low concentrations PD123319 attenuates cardiopulmonary injury by reducing pulmonary inflammation and fibrinosis and preventing PAH-induced RVH but does not affect alveolar and vascular development in newborn rats with experimental BPD.

bronchopulmonary dysplasia; angiotensin II type 2 receptor; lung inflammation; right ventricular hypertrophy; pulmonary hypertension

BRONCHOPULMONARY DYSPLASIA (BPD) is a chronic lung disease that is frequently seen in very premature infants with interrupted lung development and lung damage caused by mechanical ventilation and/or reactive oxygen species generated by (prolonged) exposure to supplemental oxygen. The hallmark of BPD is alveolar enlargement due to a permanent alveolar simplification secondary to an arrest in alveolar and vascular development, and a subsequent reduction of the alveolar surface and lung function (5, 17, 24). BPD is complicated by inflammation and oxidative stress in the neonatal period, and progressive disease ultimately results in pulmonary arterial hypertension (PAH), which is characterized by persistent vasoconstriction and structural remodeling of the pulmonary blood vessels with increased proliferation of vascular smooth muscle cells and a fixed reduction in blood vessel lumen. PAH induces right ventricular hypertrophy (RVH) and right heart failure (1, 5). PAH-induced right heart failure in children and adults is associated with a high mortality (1, 2, 37, 40). Newborn rats exposed to hyperoxia and premature infants with severe BPD develop chronic lung inflammation, persistent alveolar simplification, fibrosis, PAH, and RVH (5, 13, 14). The rodent hyperoxia model is valuable in identifying candidate interventions for BPD (4, 13, 14, 38, 47, 49).

Normal regulation of cardiovascular and renal function is affected by the renin angiotensin system and an imbalance between the opposing effector molecules angiotensin II (AngII) and angiotensin-(1–7) [Ang-(1–7)] may contribute to cardiovascular pathogenesis and lung fibrosis (19, 33, 41, 42). AngII is stepwise produced by cleavage of the prohormone angiotensinogen into the NH2-terminal decapetide AngI by renin, and the conversion of AngI to AngII by angiotensin-converting enzyme (ACE). The ligand of the Mas oncogene receptor (MAS), Ang-(1–7), is generated directly via the conversion of AngII by ACE2 or indirectly via AngI by ACE2 and ACE. AngII exerts its biological effects after binding to angiotensin II type 1 (AT1) or -type 2 receptors (AT2). Stimulation of the ACE-AngII-AT1 axis leads to vasoconstriction, proliferation, and fibrosis in multiple tissues, including the lung. Blocking AT1 or stimulation of the ACE2-Ang-(1–7)-MAS axis counterbalances the detrimental biological actions of AngII by inducing vasodilation and by inhibiting fibrinogenesis, thrombogenesis, hypertension, cardiac hypertrophy, and lung injury (8, 10, 19, 22, 25, 27, 29, 30, 32, 34, 39, 42, 45, 48). The biological response of AT1 activation by AngII may be modulated by receptor dimerization, resulting in aggravation of the response by an AT1-bradykinin type 2 heterodimer and attenuation by a dimer of AT1 and MAS oncogene receptor compared with the AngII-induced response of the AT1 homodimer (23). Alternatively, the detrimental effects of AngII binding to AT1 may be affected by binding to AT2 (36, 43). AT2 signaling can result in either beneficial or adverse effects on proliferation, inflammation, and fibrosis in cardiovascular disease (19, 36, 43), suggesting that knowledge of the role of AT2
signaling in cardiopulmonary disease is still incomplete and controversial. This may be explained by low AT2 expression in adult tissues and the complexity of AT2 signaling that can be dependent on the presence of multiple receptors, including AT1 (6). Potential adverse effects of AT2 signaling in cardiopulmonary disease are demonstrated in mice in which pharmacological blocking of AT2 attenuates bleomycin-induced fibrosis (46). Potential beneficial effects of AT2 signaling in cardiopulmonary disease are demonstrated in AT2-deficient mice that suffer from aggravated acute lung and heart injury compared with wild-type controls (3, 15) and by our recent data that demonstrated the therapeutic potential of an AT2 agonist in newborn rats with experimental BPD (45). The newborn rat is an excellent model to study AT2-dependent signaling in cardiopulmonary disease because of the 100-fold higher level of expression of AT2 in neonatal lung compared with adults (45).

To provide mechanistic data that PD123319 acts on the AT2 receptor by blocking activation of AT2 receptors in neonatal cardiopulmonary disease in vivo we investigated the role of AT2 inhibition with PD123319 (0.5 and 2 mg·kg⁻¹·day⁻¹) on the beneficial effect of AT2 agonist LP2–3 (5 µg/kg twice a day) on RVH in newborn rats with hyperoxia-induced BPD. After finding beneficial effects of treatment with the AT2 antagonist PD123319 (0.1–5 mg·kg⁻¹·day⁻¹) on RVH in a pilot experiment we determined the cardiopulmonary effects of daily treatment with the optimal dose of PD123319 (0.1 mg·kg⁻¹·day⁻¹) in the absence of the AT2 agonist in two animal models of hyperoxia-induced experimental BPD: 1) daily treatment during exposure to 100% oxygen for 10 days and 2) treatment starting on day 6 in rat pups exposed postnatally to hyperoxia for 9 days, followed by a 9-day recovery period in room air (RA).

MATERIALS AND METHODS

Animals

The research protocol was approved by the Institutional Animal Care and Use Committee of the Leiden University Medical Center. Adult Wistar rats (6 mo old; N = 8) were exsanguinated after induction of anesthesia with an intraperitoneal injection of ketamine (50 mg/kg) and xylazine (50 mg/kg). Organs were stored at −80°C until isolation of RNA for real-time RT-PCR.

For each experiment, newborn rat pups from two to three litters were pooled and distributed over the experimental group. For the intervention experiments, newborn rat pups were distributed over two experimental groups (N = 12), i.e., an oxygen and an oxygen-intervention (PD123319 and/or LP2–3) group, and two RA-exposed control groups (N = 6) injected with either saline or PD123319. Pups were fed by foster dams. Foster dams were rotated daily between the oxygen-exposed pups and two groups of RA-exposed pups to avoid oxygen toxicity: 24 h in 100% oxygen and 48 h in RA. Oxygen concentration, body weight, evidence of disease, and mortality were monitored daily.

Early concurrent treatment. Pups were continuously exposed to 100% oxygen for 10 days from birth onward. Starting on day 2 pups received daily subcutaneous injections of either the optimal concentration of 0.1 mg·kg⁻¹·day⁻¹ (135 nmol·kg⁻¹·day⁻¹) of AT2 antagonist PD123319 (1276, Axon Medchem, Groningen, The Netherlands) in 100 µl of 0.9% saline or 100 µl of 0.9% saline (age-matched control). Lung and heart tissue was collected on day 10.

To find the optimal dosing of PD123319 we performed two pilot experiments: 1) hyperoxia-exposed rat pups were treated with 0.5 or 2 mg·kg⁻¹·day⁻¹ PD123319 and/or the optimal dose of LP2–3, 5 µg/kg twice a day (a gift from Lanthio Pharma, Groningen, The Netherlands; Ref. 45) or saline (N = 8) and 2) hyperoxia-exposed rat pups were treated with 0.1–5 mg·kg⁻¹·day⁻¹ PD123319 or saline (N = 10). Because PD123319 and LP2–3 have a similar half-life in vivo in adult rats of ~20 min (Ref. 7 and Lanthio Pharma, unpublished results), 2 mg/kg of PD123319 will result in a 480-fold molar excess compared with 5 µg/kg of LP2–3 and is therefore expected to block AT2 in the presence of LP2–3, provided that both AT2 interacting compounds are simultaneously administered. We used the ratio of right and left free ventricular wall thickness (RV/LV) in histological sections of the heart as a readout. This parameter was selected for two reasons: 1) hyperoxia-induced experimental BPD results in persistent PAH-induced RVH (13, 14) and 2) stimulation of AT2 reduces PAH-induced RVH (45). Pups exposed to hyperoxia developed RVH, which could be completely prevented by administration of PD123319 (0.1 mg·kg⁻¹·day⁻¹; Fig. 1B). This concentration of PD123319 was then used in separate experiments for 1) histology (N = 10), 2) lung tissue homogenates (N = 10), 3) bronchoalveolar lavage fluid (BALF, N = 12), and measurement of chemokine-induced neutrophilic chemotactant-1 (CINC1) in BALF (N = 10). To quantify the degree of RVH, hearts were harvested, followed by removal of the atria. Next, the right ventricular (RV) free wall was dissected, weighed separately from the interventricular septum (IVS) and left ventricle (LV), frozen immediately in liquid nitrogen, and stored at −80°C for RNA isolation. The weight ratio RV/(LV + IVS) was calculated as an indicator of RVH (N = 10). In a separate experiment tissues were collected from 6-mo-old adult rats for RT-PCR (N = 8).

Late treatment and recovery. Lung injury and recovery were investigated by exposing pups to hyperoxia for 9 days, followed by recovery in RA for 9 days. After 6 days of hyperoxia, daily injections with 0.9% saline or 0.1 mg·kg⁻¹·day⁻¹ of PD123319 in 0.9% saline were started and continued throughout the 9-day recovery period in RA. Lung and heart tissue was collected on day 9, after 9 days of hypoxic lung injury (N = 8), and on day 18, after 9 days of recovery in RA (N = 8).

Histology

Pups were anesthetized with an intraperitoneal injection of ketamine (25 mg/kg body wt; Nematek, Eurovet Animal Health BV, Bladel, The Netherlands), xylazine (50 mg/kg body wt; Rompun, Bayer, Leverkusen, Germany), and heparin (100 units; Leo Pharma, Breda, The Netherlands, to avoid postmortem fibrin deposition in the lungs) on day 10. After 5 min, pups were exsanguinated by transection of the abdominal blood vessels and the trachea was cannulated (Bioflow 0.6-mm intravenous catheter, Vygon, Veenendaal, The Netherlands) for perfusion fixation of the lungs with buffered formaldehyde (4% paraformaldehyde in PBS, pH 7.4) at 25 cm H₂O pressure for 6 min. 15 min after bleeding the heart did not beat anymore when the thorax was opened and lungs and heart were removed. The heart was submerged in cold 0.9% NaCl for cardioplegia, as the heart stops beating and remains in diastole. Hereafter heart and lungs were fixed (additionally) in cold formaldehyde for 24 h at 4°C and embedded in paraffin after dehydration in a graded alcohol series and xylene. Formalin-fixed, paraffin-embedded, 4-µm-thick heart and lung sections were stained with hematoxylin and eosin. Immunohistochemistry was performed using the chromogenic substrate NovaRed as recommended by the manufacturer (Vector, Burlingame, CA), and counterstained...
brieﬂy with hematoxylin by standard methods (14, 45). Extracellular collagen III deposition was quantiﬁed on lung sections stained immunohistochemically for collagen III at a 400-fold magniﬁcation in 10 nonoverlapping ﬁelds. Morphometric quantiﬁcation of alveolar enlargement, vascularization, septal thickness, the pulmonary inﬁltrate of neutrophils and macrophages, and right ventricular hypertrophy was performed by two independent researchers blinded to the treatment strategy as previously described (20, 45, 49; N = 8).

Fibrin Detection Assay

Quantitative ﬁbrin deposition in lung tissue homogenates was determined by Western blotting, using the monoclonal 5D8 (Oklahoma Medical Research Foundation, Oklahoma City, OK; diluted 1:1,000) and an infrared detection system (Odyssy infrared imaging system, Licor Biosciences) as described previously (44, 45; N = 10).

Bronchoalveolar Lavages, Protein Assay, and Cytokine Measurement

Total protein content in BALF was determined as an indicator of vascular leakage using a standard protein assay (DC Protein Assay, Measurement Systems, Licor Biosciences) as described previously (44, 45; N = 10). In BALF CINC1 protein was measured with a commercially available ELISA (catalog no. ADI-900-074, Enzo Life Sciences, Raamsdonksveer, The Netherlands), as recommended by the manufacturer.

Real-time RT-PCR

Total RNA isolation from lung tissue homogenates (RNA-Be, Tel-Test, Bio-Connect, Huissen, The Netherlands), ﬁrst-strand cDNA synthesis [SuperScript Choice System (Life Technologies, Breda, The Netherlands), ﬁrst-strand cDNA production (Tel-Test, Bio-Connect, Huissen, The Netherlands), as recommended by the manufacturer.

Table 1. Sequences of oligonucleotides for forward and reverse primers for real-time RT-PCR

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<th>Gene Product</th>
<th>Forward Primer</th>
<th>Reverse Primer</th>
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<td>5’-AGGAAAGAAAGAAATGATC-3’</td>
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Fig. 1. Effect of PD123319 (0.5 and 2 mg·kg⁻¹·day⁻¹) on angiotensin receptor type 2 (AT2) agonist (LP2–3: 5 µg/kg twice a day)-induced prevention of hyperoxia-induced right ventricular hypertrophy (RVH) in newborn rat pups on day 10 after birth by morphometry in parafﬁn sections stained with hematoxylin and eosin and depicted as right ventricle (RV)/left ventricle (LV) wall thickness ratio (N = 6; A). Pilot experiment to ﬁnd the optimal dose of PD123319 for treatment of experimental bronchopulmonary dysplasia (BPD) by determining RVH depicted as RV/LV wall thickness ratio on day 10 after birth of rat pups (N = 10; B). Rat pups were exposed to room air (RA) and injected daily with saline (open bars) or of PD123319: 0.1, 0.5, 2, and 5 mg·kg⁻¹·day⁻¹ and/or LP2–3 (shaded bars). Data are expressed as means ± SE. *P < 0.05 and **P < 0.001 vs. RA controls. $P < 0.05 vs. PD123319 (0.5 mg·kg⁻¹·day⁻¹).

Statistical Analysis

Values are expressed as means ± SE. Differences between groups were analyzed by one-way ANOVA for independent samples, followed by Tukey’s multiple comparison test using the GraphPad Prism, version 5, software package (San Diego, CA). Differences at P values < 0.05 were considered statistically signiﬁcant.
RESULTS

Dose Finding and Effects of PD123319 on AT2-Agonist-Induced Prevention of Right Ventricular Hypertrophy

Development of RVH by exposure of newborn rats to hyperoxia for 10 days, shown by an increase (1.4-fold, Fig. 1A; 1.3-fold, Fig. 1B) in the RV/LV free wall thickness ratio over RA controls, was prevented by twice daily administration of 5 μg/kg of AT2 agonist LP2–3 (Fig. 1A and Ref. 45). This beneficial effect of LP2–3 on RVH was abolished by coadministration of PD123319 (0.5 and 2 mg·kg·day⁻¹). Administration of 0.5 mg·kg·day⁻¹ of PD123319 had no significant effect on hyperoxia-induced RVH. Because 0.5 mg·kg·day⁻¹ of PD123319 showed a tendency toward lower levels of RVH (P = 0.06; Fig. 1A) we subsequently investigated the beneficial effect of PD123319 on RVH. Administration of 0.1 mg·kg·day⁻¹ of PD123319 for 10 days was the most optimal concentration to prevent RVH in hyperoxia-induced BPD, demonstrated by a normalization in relative RV/LV free wall thickness (P < 0.001; Fig. 1B).

Effects of PD123319 on Growth and Survival

Early concurrent treatment. On day 10, mean body weight of pups was comparable in all RA groups (19 g; Fig. 2A) and all oxygen groups (12–13 g). Administration of PD123319 (0.1 mg·kg·day⁻¹) for 10 days had no adverse effect on mean body weight in RA controls and oxygen-exposed pups. Exposure to hyperoxia resulted in a 70% survival on day 10 and was not affected by administration of PD123319 (Fig. 2B). RA-exposed pups showed no morbidity or mortality during the experimental period of 10 days.

Late treatment and recovery. Mean body weight of RA controls was 18.2 g on day 9 (Fig. 2C) and 37.6 g on day 18 and was not influenced by PD123319 treatment. After 9 days of hyperoxia exposure, mean body weight was 11.5 g and this increased to 24.5 g after 9 days of recovery in RA on day 18. Treatment with PD123319 did not affect body weight compared with oxygen-exposed controls on days 9 and 18. On days 9 and 18 all RA controls survived (Fig. 2D), but exposure to hyperoxia resulted in a 70% survival. Treatment with PD123319 for 3 days did not affect survival on day 9, and 80% of the pups that recovered in RA survived until day 18.

Fig. 2. Growth (A and C) and survival (B and D) on day 10 of prophylactic PD123319 treatment (N = 12; A and B) and after late treatment and recovery (N = 8; C and D) on days 9 and 18 in RA and age-matched O₂-exposed pups injected daily with saline PD123319 (0.1 mg·kg·day⁻¹). Data are expressed as means ± SE. Open bars, RA-NaCl; hatched bars, RA-PD123319; solid bars, O₂-NaCl; shaded bars, O₂-PD123319. Data are expressed as means ± SE. *P < 0.05 and ***P < 0.001 vs. age-matched O₂-exposed controls. ΔΔΔP < 0.001 vs. own RA-exposed controls. &P < 0.001 vs. own treatment controls on day 9.

Effects of PD123319 on Lung Airway Development and Inflammation

Early concurrent treatment. In the rat lung development proceeds from the saccular stage at birth toward the alveolar stage in 10 days (Fig. 3A). Administration of PD123319 (0.1 mg·kg·day⁻¹) did not have adverse effects on the number of alveolar crests (Fig. 4A), pulmonary vessel density (Figs. 3B and 4B), alveolar septal thickness (Figs. 3B and 4C), arterial medial wall thickness (Figs. 3F and 4D), and influx of macrophages (Figs. 3J and 4E) and neutrophilic granulocytes (Figs. 3N and 4F). Oxygen exposure for 10 days resulted in edema,
a heterogeneous distribution of enlarged air spaces with a decreased number of alveolar crests (2.1-fold, \( P < 0.001 \); Fig. 4A), surrounded by septa with increased thickness (1.6-fold, \( P < 0.001 \); Figs. 3C and 4C), reduced pulmonary vessel density (2.7-fold, \( P < 0.001 \); Figs. 3C and 4B), and increased pulmonary arterial medial wall thickness (2.3-fold, \( P < 0.001 \); Figs. 3G and 4D). Hyperoxia led to a massive inflammatory reaction, characterized by an overwhelming influx of inflammatory cells, including macrophages (5.4-fold, \( P < 0.001 \); Figs. 3K and 4E) and neutrophils (2.8-fold, \( P < 0.001 \); Figs. 3O and 4F), compared with RA-exposed controls. Administration of PD123319 reduced alveolar septal thickness by 17% (\( P < 0.001 \); Figs. 3D and 4C), arterial medial wall thickness by 25% (\( P < 0.01 \); Figs. 3H and 4D), and the influx of macrophages by 29% (\( P < 0.01 \); Figs. 3L and 4E) and neutrophils by 54% (\( P < 0.001 \); Figs. 3P and 4F) compared with oxygen-exposed controls but had no beneficial effects on hyperoxia-induced inhibition of alveolarization and angiogenesis.

**Late treatment and recovery.** Treatment of RA-exposed pups with PD123319 had no adverse effect on alveolar (Fig. 6A) and vascular development (Figs. 5B and 6, B and C) on days 9 (Fig. 5F) and 18 (Fig. 5F). Continuous neonatal exposure to hyperoxia for 9 days resulted in enlarged alveoli (Fig. 5C), demonstrated by a 2.3-fold decrease in the number of alveolar crests (\( P < 0.001 \); Fig. 6A), disturbed vascular development, demonstrated by a 2.4-fold reduction in blood vessel density (\( P < 0.001 \); Figs. 5C and 6B), and a 2.2-fold increase in arterial medial wall thickness (\( P < 0.001 \); Figs. 5K and 6C) compared with RA controls. PD123319 treatment during the last 3 days of the injurious 9-day hyperoxic period decreased arterial medial wall thickness by 27% (\( P < 0.05 \); Figs. 5L and 6C) but had no beneficial effects on alveolarization (Figs. 5D and 6A) and pulmonary blood vessel density (Figs. 5D and 6B). A recovery period of 9 days in RA after hyperoxia-induced lung injury on day 18 had a beneficial effect on the number of alveolar crests (\( P < 0.001 \); Fig. 6A), blood vessel density (\( P < 0.001 \); Fig. 6B) and arterial medial wall thickness (\( P < 0.05 \); Fig. 6C), but the number of alveoli and blood vessels were still reduced and arterial medial wall thickness was still increased after injury and recovery. Treatment with PD123319 reduced arterial medial wall thickness by 32% (\( P < 0.05 \); Figs. 5P and 6C) but did not have a beneficial effect on alveolarization (Fig. 6).

**Fig. 3.** Representative lung sections stained for von Willebrand factor (vWF; A–D), \( \alpha \)-smooth muscle actin (ASMA; E–H), the monocyte and macrophage marker ED1 (I–L), or myeloperoxidase (MPO) as a marker for neutrophilic granulocytes (M–P) of RA (A, B, E, F, I, J, M, and N) and O2-exposed pups (C, D, G, H, K, L, O, and P) injected daily with saline (A, C, E, G, I, K, M, and O) or PD123319 (PD; 0.1 mg·kg\(^{-1}\)·day\(^{-1}\); B, D, F, H, J, L, N, and P) until 10 days of age. a, Alveolus. Arrows in A–D indicate vWF-positive blood vessels.
and vascularization (Figs. 5H and 6B), compared with nontreated experimental BPD pups at the end of the recovery period on day 18.

Effects of PD123319 on Lung Coagulation, Collagen Deposition, Vascular Leakage, and CINC1 Expression

Early concurrent treatment. Collagen III was present at high levels in the perivasculature of large and small blood vessels in normal lung in the absence (Fig. 7A) or presence of PD123319 (Fig. 7B). In alveolar septa expression was low or absent. In lungs of pups exposed to hyperoxia for 10 days, collagen III deposition increased 6.3-fold ($P < 0.001$; Fig. 7E) and was present in the perivasculature of blood vessels and in thick alveolar septa (Fig. 7C). Treatment with PD123319 for 10 days reduced collagen III expression by 52% ($P < 0.001$; Fig. 7E). Hyperoxia-induced extracellular collagen III expression was only reduced in alveolar septa, but not in the (peri)vascular area (Fig. 7D). Fibrin deposition was at reference levels during normal neonatal pulmonary development on day 10 in the absence or presence of PD123319 ($< 15$ ng fibrin/mg tissue) and increased 11-fold ($P < 0.01$) in lungs of pups exposed to 100% oxygen for 10 days (Fig. 7F). PD123319 (0.1 mg·kg$^{-1}$·day$^{-1}$) administration did not reduce hyperoxia-induced fibrin deposition. Protein expression of CINC1 was low in BALF of RA controls and increased 6.4-fold ($P < 0.05$; Fig. 7G) after exposure to hyperoxia for 10 days. Daily administration of 0.1 mg/kg of PD123319 showed a tendency toward lower CINC1 levels. Total protein concentration in BALF was determined to establish the inhibitory effect of PD123319 on pulmonary edema by capillary-alveolar leakage (Fig. 7H). The protein concentration on postnatal day 10 increased 5.3-fold after hyperoxia ($P < 0.05$) and was not affected by PD123319 administration during normoxia or hyperoxia.

Effects of PD123319 on mRNA Expression in Lung Tissue

Early concurrent treatment. Administration of PD123319 (0.1 mg·kg$^{-1}$·day$^{-1}$) for 10 days during normal neonatal development in RA did not change mRNA expression (Fig. 8) of the proinflammatory factors monocyte chemotactic protein (MCP)-1 (Fig. 8A) and CINC1 (Fig. 8B), the anti-fibrinolytic protein plasminogen activator inhibitor 1 (PAI-1; Fig. 8C), angiotensin II type 1a receptor (AT1a; Fig. 8D), AT2 (Fig. 8E), MAS (Fig. 8F), ACE (Fig. 8G), and ACE2 (Fig. 8H). Ten days of oxygen exposure resulted in an increase in mRNA expression of MCP-1 (5.4-fold, $P < 0.001$), CINC-1 (12.1-fold, $P < 0.001$), PAI-1 (53-fold, $P < 0.001$), and AT2 (4.1-fold, $P < 0.001$), whereas mRNA expression was decreased in lungs of oxygen-exposed pups for AT1a (1.8-fold, $P < 0.001$), MAS (1.6-fold, $P < 0.001$), and ACE (4.2-fold, $P < 0.001$) compared with RA controls. Treatment of oxygen-exposed pups with PD123319 for 10 days did not result in changes in mRNA expression compared with oxygen-exposed pups.

Tissue Distribution of MAS-Related Receptor Type D

Because PD123319 (0.1 mg·kg$^{-1}$·day$^{-1}$) may exert its biological effects by inhibiting AT2 and MAS-related receptor type D (MrgD; Ref. 21), we studied the tissue distribution of MrgD and found that MrgD is expressed in testis, but not in the other organs studied, including lung and heart (Fig. 8I and Ref. 35). In addition, MrgD expression was absent in lungs of PD123319-treated rat pups with experimental BPD and controls (data not shown).
Effects of PD123319 on Right Ventricular Hypertrophy

Early concurrent treatment. Administration of 0.1 mg·kg\(^{-1}\)·day\(^{-1}\) of PD123319 for 10 days during normal neonatal development had no adverse effect on the heart (Fig. 9, A–C). Exposure to hyperoxia for 10 days resulted in RVH, affected by an increase (1.3-fold; Fig. 9B) in the ratio RV/LV free wall thickness and an 1.4-fold increase in RV/(LV + IVS) weight ratio (Fig. 9C) compared with RA controls (P < 0.01). Administration of 0.1 mg·kg\(^{-1}\)·day\(^{-1}\) of PD123319 prevented RVH, demonstrated by a normal-

Fig. 5. Representative lung sections stained for vWF (A–H) or ASMA (I–P) after late treatment and recovery on days 9 (A–D and I–L) and 18 (E–H and M–P) of RA (A, B, E, F, I, J, M, and N) and O\(_2\)-exposed pups (C, D, G, H, K, L, O, and P) injected daily with saline (A, C, E, G, I, K, M, and O) or PD123319 (0.1 mg·kg\(^{-1}\)·day\(^{-1}\); B, D, F, H, J, L, N, and P) from days 6 to 18. a, Alveolus. Arrows in A–H indicate vWF-positive blood vessels.

Fig. 6. Quantification of alveolar crest (A), number of pulmonary vessels (B), and arterial medial wall thickness (C) determined on paraffin sections after late treatment and recovery on days 9 and 18 in RA-exposed pups injected daily with saline (open bars) or PD123319 (hatched bars) and O\(_2\)-exposed pups injected daily with saline (solid bars) or PD123319 (shaded bars). Values are expressed as means ± SE (N = 8). *P < 0.05 and **P < 0.001 vs. age-matched O\(_2\)-exposed controls. \(\Delta\Delta\Delta\)P < 0.01 and \(\Delta\Delta\Delta\)P < 0.001 vs. own RA controls. \(\delta\)P < 0.05 and \(\delta\delta\delta\)P < 0.001 vs. own treatment controls on day 9.
as a marker for lung injury; CINC1 protein expression in bronchoalveolar lavage fluid (BALF; as a marker for vascular leakage on saline (A; Fig. 7. Quantification of collagen III (Col3) expression (E) on lung paraffin sections of RA (A and B) and O2-exposed pups (O2; C and D) injected daily with saline (A and C) or PD123319 (PD; B; D) until 10 days of age (N = 8); quantification of extravascular fibrin deposition in lung homogenates (N = 10, F) as a marker for lung injury; CINC1 protein expression in bronchoalveolar lavage fluid (BALF; N = 10, G); and total protein concentration in BALF (N = 12, H) as a marker for vascular leakage on day 10 in RA pups injected daily with saline (open bars) or PD123319 (0.1 mg·kg−1·day−1; hatched bars) and O2 pups injected daily with saline (solid bars) or PD123319 (shaded bars). a. Alveolus; at. arteriole. Values are expressed as means ± SE. *P < 0.05, **P < 0.01, and ***P < 0.001 vs. age-matched O2-exposed controls. ΔP < 0.05 and ΔΔP < 0.01 vs. RA controls.

**Discussion**

PAH and RVH are major causes of mortality and severe morbidity in premature infants with severe BPD (1, 5). The hyperoxia-exposed newborn rat pup is an established in vivo model for severe experimental BPD and RVH (44, 45). Because of the high expression of AT2 in neonatal rat lung (45), neonatal rats with BPD are very suitable to study novel treatment options that target AT2. Treatment of neonatal rats with experimental BPD with the AT2 agonist LP2–3 attenuated hyperoxia-induced BPD. The relatively high expression of AT2 and the adaptive mRNA response in the newborn lung exposed to hyperoxia, result in a higher AT2 than AT1 expression (45), and may contribute to the cardiopulmonary effects that we observed after treatment of rat pups with experimental BPD with PD123319 (this study) or the AT2 agonist LP2–3 (45).

Inflammation contributes significantly to lung injury in experimental BPD as demonstrated by 1) the massive influx of macrophages and neutrophils into the lung, 2) the upregulation of chemokines that are involved in the activation and migration of leukocytes in vivo (44, 49) and 3) the beneficial effects of PDE-4 inhibitors in experimental BPD, which are potent anti-inflammatory agents (14, 47). The reduced influx of monocytes, macrophages, and neutrophilic granulocytes in the lung suggests that the beneficial effects of treatment with PD123319 on hyperoxia-induced neonatal lung injury may be explained by reduced inflammation, but this was not confirmed by reduced mRNA expression of proinflammatory cytokines (MCP1 and CINC1), suggesting regulation of gene expression at a posttranscriptional level. A role for the AT2 receptor in the
regulation of the inflammatory response in vivo is further supported by the anti-inflammatory and/or antifibrotic effects of AT2 stimulation in neonatal lung disease (45), bleomycin-induced cutaneous injury (31), and renovascular hypertension and inflammation (26), of AT2 inhibition in bleomycin-induced lung fibrosis (46) and the aggravation of acute lung disease in AT2-deficient mice (15).

Treatment of hyperoxia-induced neonatal lung injury with PD123319 results in a significant improvement of severe cardiopulmonary injury, demonstrated by prevention of PAH and RVH and reduction of inflammation, but the beneficial effects of PD123319 are probably still too small to result in reduced mortality. The data demonstrate that the development of PAH and RVH can be attenuated by prophylactic PD123319 administration in rat pups with experimental BPD, in agreement with the beneficial effects observed in PD123319-treated mice with bleomycin-induced fibrosis (46), but in sharp contrast with our previous findings with the AT2 agonist LP2–3 in the same model (45) and observations in AT2-deficient mice in which cardiovascular disease is aggravated compared with wild-type controls (3, 15). Secondly, in previous studies we observed that beneficial effects of treatment with MAS, AT2, and APJ receptor-specific agonists cAng I–7, LP2–3, and apeelin, respectively, were only found at low concentrations in

Although the beneficial effects of MAS stimulation on cardiovascular disease in adult and neonatal animal models in the literature are rather consistent, the role of AT2 continues to be controversial as demonstrated by two observations in our study: 1) beneficial effects of the AT2 antagonist PD123319 on RVH observed in this study were also observed for treatment with AT2 agonist LP2–3 in the same model (45) and 2) these beneficial effects disappear when the PD123319 dose is increased. These unexpected finding are supported by two observations by us and others: Firstly, the beneficial effects of PD123319 administration in rat pups with experimental BPD are in agreement with the beneficial effects observed in PD123319-treated mice with bleomycin-induced fibrosis (46), but in sharp contrast with our previous findings with the AT2 agonist LP2–3 (45) and observations in AT2-deficient mice in which cardiovascular disease is aggravated compared with wild-type controls (3, 15). Secondly, in previous studies we observed that beneficial effects of treatment with MAS, AT2, and APJ receptor-specific agonists cAng I–7, LP2–3, and apeelin, respectively, were only found at low concentrations in
newborn rats with BPD and disappeared when the dose was increased (13, 45). These discrepancies may be explained by the presence of AT2 on multiple cell types, including endothelial and vascular smooth muscle cells, and may be complicated by its increased expression under pathological conditions compared with healthy controls. We speculate that AT2 activation on the endothelium may lead to vasodilation, whereas AT2 activation on vascular smooth muscle cells may lead to vasocostriction (43). In the cardiovascular system this behavior is not unique to AT2 but is also observed for other receptors related to the renin angiotensin system, including the apelin receptor API, bradykinin B1 and B2 receptors, and muscarinic receptors, and may be complicated by receptor expression on inflammatory cells and cardiomyocytes (9, 12, 16, 18, 28, 43). This makes it difficult to predict the biological effect of an AT2 agonist or antagonist, because it depends not only on the administered dose and diffusion into the tissue, but also on local levels of AngII and its intermediates and on the differential cellular expression of AT1, AT2, and MAS. Alternatively, these discrepancies may be explained by a lack of specificity of PD123319 for the AT2 receptor, as was demonstrated recently by 1) an inhibitory effect of PD123319 on the alamandine receptor MrgD (21) and 2) an increase of AngII-induced abdominal aortic aneurysms by PD123319 in AT2-deficient mice (11), possibly due to MrgD inhibition (21). However, MrgD expression is absent in lung and heart tissue (this study and Ref. 33). It is therefore unlikely that inhibition of MrgD by PD123319 is involved in the beneficial cardiopulmonary effects in rat pups with BPD, but binding of PD123319 to other proteins than AT2 or MrgD cannot be excluded. In addition, PD123319 may act as an AT2 agonist at low concentrations and mimic the beneficial effects of the AT2 agonist LP2–3 in our previous study (45). Also pleiotropic effects of LP2–3 will have to be investigated. Off-target effects of both protective compounds would offer possibilities to discover new therapeutic targets.

Fig. 9. RVH was determined by morphometry in paraffin sections stained with hematoxylin and eosin (A) and depicted as RV/LV wall thickness ratio (B and D) or by determining the weight ratio RV/(LV + IVS) where IVS is interventricular septum (C) on day 10 of prophylactic PD123319 treatment (A–C) and after late treatment and recovery (D) on days 9 and 18 in RA and age-matched O2-exposed pups injected daily with saline or PD123319 (0.1 mg·kg⁻¹·day⁻¹). White bars, RA-NaCl; hatched bars, RA-PD123319; solid bars, O2-NaCl; shaded bars, O2-PD123319. Data are expressed as means ± SE (N = 8). *p < 0.05, **p < 0.01, and ***p < 0.001 vs. own age-matched O2-exposed controls. \( \Delta p < 0.05 \) vs. own RA controls.
showed additional beneficial effects on reduced alveolarization and vascularization. However, prolonged sildenafil treatment of children with pulmonary arterial hypertension resulted in increased mortality and let the FDA to give a safety warning against the use (particularly chronic use) of sildenafil in children.

Extrapolation of our experimental findings to the clinic would suggest a beneficial effect of PD123319 on inflammation, fibrosis, PAH, and RVH and the possibility of combating these major causes of mortality or severe morbidity in premature infants with severe BPD.

ACKNOWLEDGMENTS

The authors gratefully acknowledge I. van Ark and T. A. Leusink-Muis (Department of Pharmacology, Utrecht University, Utrecht, The Netherlands) for expert technical assistance, Dr. E. de Heer (Department of Pathology, Leiden University Medical Center, Leiden, The Netherlands) for providing the ED-1 antibody, and Dr. J. J. Baelde (Department of Pathology, Leiden University Medical Center, Leiden, The Netherlands) for providing the COL3A antibody.

GRANTS

This study was supported by grants IR01 HL092158 and IR01 ES015330 from the National Institutes of Health (F. J. Walther), Dutch Heart Foundation research grant NHS2010B0009 (A. J. M. Roks), and a grant from the Stichting Zeldzame Ziekten Fonds (G. T. M. Wagenaar).

DISCLOSURES

No conflicts of interest, financial or otherwise, are declared by the author(s).

AUTHOR CONTRIBUTIONS


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