C-peptide, a Novel Inhibitor of Lung Inflammation following Hemorrhagic Shock

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ABSTRACT

C-peptide is a 31-amino acid peptide cleaved form proinsulin during insulin synthesis. Initially thought to be inert, recent work suggests that it may modulate the inflammatory response in the setting of endotoxemia and ischemia reperfusion. However, the spectrum of its biologic effects is unclear. We hypothesized that exogenous administration of C-peptide would modulate pro- and anti-inflammatory signaling pathways and thereby attenuate lung inflammation in an in vivo model of hemorrhagic shock. Hemorrhagic shock was induced in male wistar rats (aged 3-4 months) by withdrawing blood to a mean arterial pressure (MAP) of 50 mmHg. At 3 hours after hemorrhage, rats were rapidly resuscitated by returning their shed blood. At the time of resuscitation and every hour thereafter, animals received C-peptide (280 nmol/kg) or vehicle parenterally. Animals were euthanized at 1h and 3h after resuscitation. C-peptide administration at resuscitation following hemorrhagic shock ameliorated hypotension and blunted the systemic inflammatory response by reducing plasma levels of interleukin-1 (IL-1), interleukin-6 (IL-6), macrophage inflammatory protein-1 alpha (MIP-1α) and cytokine-induced neutrophil chemoattractant-1 (CINC-1). This was associated with a reduction in lung neutrophil infiltration and plasma levels of receptor for advanced glycation end-products (RAGE). Mechanistically, C-peptide treatment was associated with reduced expression of pro-inflammatory transcription factors activator protein-1 (AP-1) and nuclear factor-κB (NF-κB) and activation of the anti-inflammatory transcription factor peroxisome proliferator-activated receptor-γ (PPARγ). Our data suggest that C-peptide ameliorates the inflammatory response and lung inflammation following hemorrhagic shock. These effects may be modulated by altering the balance between pro- and anti-inflammatory signaling in the lung.

Keywords: C-peptide; hemorrhage; acute lung injury; peroxisome proliferator-activated receptor-γ
INTRODUCTION

Hemorrhagic shock and resuscitation is associated with a development of an inflammatory state which may lead to the development of multiple organ failure and death (14, 15). Lung injury is a common occurrence and may contribute to other distant organ failures (14). Clinically the diagnosis of acute lung injury (ALI) is based on consensus criteria which do not account for the underlying etiology of lung injury (9). Recent studies in rodents and humans have assessed the utility of biomarkers for diagnosing ALI and demonstrated the potential role of plasma levels of the multi-ligand binding receptor RAGE as a marker of lung injury (17, 33).

At a molecular level, lung injury and systemic inflammation following hemorrhagic shock and resuscitation is characterized by a complex process that involves both pro-inflammatory and anti-inflammatory signaling pathways and is characterized by dysregulated production of pro-inflammatory enzymes, cytokines and adhesion molecules. The pro-inflammatory response is characterized by activation of mitogen activated protein kinases (MAPK) such as extracellular signal-regulated kinase (ERK 1/2) and c-Jun N-terminal kinase (JNK) and IκB kinases (IKK) which mediate the transduction of oxidant stress signals to the nuclear transcription factors activator protein (AP)-1 and nuclear factor-κB (NF-κB) (11, 12, 16). The anti-inflammatory response may also be modulated by signaling pathways that involve peroxisome proliferator-activated receptor gamma (PPARγ). PPARγ is a nuclear transcription factor that regulates expression of inflammatory response genes and has been demonstrated to exert anti-inflammatory properties in models of sepsis (22, 27). Previously, in a rodent model of hemorrhagic shock we have demonstrated that ciglitazone; a PPARγ ligand ameliorated lung inflammation and reduced systemic levels of cytokines and chemokines by modulating NF-κB signaling (12). Hence, using a strategy aimed at down regulating pro-inflammatory pathways and simultaneously up-regulating the anti-inflammatory pathways may attenuate the inflammatory response following hemorrhagic shock.
C-peptide is a 31-amino acid peptide cleaved from proinsulin during insulin synthesis (31). It is stored and released into the circulation from the pancreatic β cells in equimolar concentrations with insulin. C-peptide was thought to have minimal biological activity; however recent data suggest that this may not be the case (35). Specifically, in diabetic animals it attenuates neural and vascular dysfunction, while in non-diabetic animals it may mediate leukocyte endothelial interactions; modulate nitric oxide (NO) production and neutrophil infiltration (20, 29, 37). The details of its putative receptor and its cellular mechanisms of action are unclear; however, it appears that C-peptide may exert its biologic effects by binding to a G protein coupled receptor (28, 35). Recently published data indicates that C-peptide may attenuate lung injury in a non-diabetic rodent model of sepsis as a result of activation of PPARγ and modulation of MAPK signaling (5, 34). Therefore, C-peptide may act as an endogenous factor modulating both pro-inflammatory and anti-inflammatory signaling pathways. However, the biological effects of exogenous administration of C-peptide on inflammation following hemorrhagic shock are unknown.

Our current study aimed at exploring biological effects of C-peptide on the systemic inflammatory response and lung inflammation following severe hemorrhage in an in vivo rodent model of hemorrhagic shock. We hypothesized that exogenous administration of C-peptide would be beneficial by modulating pro- and anti-inflammatory signaling pathways.
MATERIAL AND METHODS

Rodent Model of Hemorrhagic Shock and Resuscitation (HS/R)

Male Wistar rats aged 3-4 months (Charles River Laboratories, Wilmington MA) were anesthetized with pentobarbital (80 mg/kg) intraperitoneally (IP). The trachea was cannulated and the animals were placed on a rodent ventilator (Harvard Apparatus, Holliston, MA) with similar settings, tidal volume 2 ml, rate 60 breaths/min and FiO2 of 0.4. The right femoral artery and left common carotid artery were cannulated (PE-50 tubing) and used for drawing blood and measuring mean arterial blood pressure (MAP), respectively. Following the surgical procedure rats were allowed to stabilize for 10 minutes. Hemorrhagic shock was induced by withdrawing blood from the femoral artery over 10 minutes into a reservoir as previously described (42). The animals were kept at a MAP of 50 mmHg for 3h by withdrawing or re-injecting blood through the femoral artery. At 3h rats were rapidly resuscitated over 10 min with their own shed blood supplemented with Ringer Lactate solution to a final volume equal to total shed blood and monitored for another 3h. Heart rates (HR) and MAP were measured using a pressure transducer and digitized using a Maclab A/D converter. This data was analyzed using Chart 5 software (AD Instruments, Colorado Springs, CO) at 30 min intervals during the entire experiment. The investigation conformed to the Guide for the Care and Use of Laboratory Animals published by the U.S. National Institutes of Health (NIH Publication No. 85-23, revised 1996) and was approved by the institutional Animal Care and Use Committee at Cincinnati Children’s Hospital Medical Center.

Experimental groups

Rats were assigned to 6 groups: 1. Control (n=5), rats in this group underwent surgery following which they were euthanized. 2. Sham (n=6), rats in this group underwent surgery but no HS/R; they were maintained under anesthesia with pentobarbital and kept on the ventilator. 3. Sham+Vehicle (n=6), rats in this group underwent surgery but no HS/R; they were treated
with vehicle (0.1% acetic acid) intra-arterially at 3h and hourly thereafter and were euthanized at 1h (n=3) and 3h (n=3) following vehicle-treatment. 4. Sham+C-Peptide (n=6), rats in this group underwent surgery but no HS/R; they were treated with C-peptide (280 nmol/Kg or 1 mg/kg) intra-arterially at 3h and hourly thereafter and were euthanized at 1h (n=3) and 3h (n=3) following C-peptide-treatment. 5. HS/R+Vehicle (n=10), rats in this group underwent surgery followed by HS/R; after 3h they were resuscitated and treated with vehicle (0.1% acetic acid) intra-arterially at resuscitation and hourly thereafter. Rats in these groups were euthanized at 1h (n=5) and 3h (n=5) following resuscitation. 6. HS/R+C-Peptide (n=10), rats in this group underwent surgery followed by HS/R; after 3h they were resuscitated and treated with C-peptide (280 nmol/Kg or 1mg/kg) intra-arterially at resuscitation and hourly thereafter. Rats in these groups were euthanized at 1h (n=5) and 3h (n=5) following resuscitation.

The dose of C-peptide was chosen on the basis of prior work in an in vivo model of endotoxemia (34). Acetic acid was the vehicle used to reconstitute C-peptide and was used as the vehicle. Arterial whole blood (100 μl) was collected and analyzed using a hand held analyzer (iStat 1, Abbot Laboratories, IL) for lactate and glucose levels following surgery (baseline), at the end of hemorrhage and at the end of the experiment. Following euthanasia lungs and plasma samples were collected from each rat at the end of the experiment and stored at -70°C. A part of the lung tissue was collected and stored in formalin for histopathology.

**Plasma Cytokine and Chemokine Levels**

Plasma levels of interleukin 1 (IL-1), interleukin 6 (IL-6), macrophage inflammatory protein-1 alpha (MIP-1α) were determined by a multiplex array system (Millipore, Billerica, MA). Cytokine-induced neutrophil chemoattractant 1 (CINC-1) levels were determined using an ELISA technique (R&D Systems, Minneapolis, MN).
Plasma RAGE Levels

Plasma levels of RAGE were quantified using an ELISA technique as specified by the manufacturer (RayBiotech Inc, Norcross, GA).

MPO Activity

Myeloperoxidase (MPO) activity was determined as an index of lung neutrophil infiltration. It was defined as the quantity of enzyme degrading 1μmol hydrogen peroxide/min at 37ºC and expressed in units per 100 mg tissue (43).

Histopathological analysis

Lung tissue was fixed in 4% paraformaldehyde and embedded in paraffin. Sections were stained with hematoxylin eosin stain. These sections were evaluated by light microscopy for evidence of lung inflammation and injury.

Subcellular Fractionation and Nuclear Protein Extraction

Lung samples were homogenized with a Polytron homogenizer (Brinkmann Instruments, West Orange, NY) in a buffer containing 0.32 M sucrose, 10 mM Tris-HCl (pH 7.4), 1 mM EGTA, 2 mM EDTA, 5 mM NaN3, 10 mM β-mercaptoethanol, 20 μM leupeptin, 0.15 μM pepstatin A, 0.2 mM PMSF, 50 mM NaF, 1 mM sodium orthovanadate, and 0.4 nM microcystin. The homogenates were centrifuged (1,000 x g, 10 min), and the supernatant (cytosol plus membrane extract) was collected to evaluate the content of PPARγ, pERK1/2 and ERK1/2. The pellets were solubilized in Triton buffer (1% Triton X-100, 150 mM NaCl, 10 mM Tris-HCl (pH 7.4), 1 mM EGTA, 1 mM EDTA, 0.2 mM sodium orthovanadate, 20 μM leupeptin A, and 0.2 mM
PMSF). The lysates were centrifuged (15,000 x g, 30 min, 4°C), and the supernatant (nuclear extract) was collected.

**Western Blot Analyses**

The content of PPAR\(\gamma\), pERK and ERK were determined by immunoblot analyses using primary antibody against PPAR\(\gamma\), pERK and ERK. Immunoreaction was visualized by chemiluminescence and on a photographic film. Densitometric analysis of blots was performed using ImageQuant (Molecular Dynamics, Sunnyvale, CA).

**AP-1 and PPAR\(\gamma\) Activation**

AP-1 and PPAR\(\gamma\) Activation was assessed by electrophoretic mobility shift assays (EMSA) as previously described (41). Oligonucleotide probes corresponding to the AP-1 or PPAR\(\gamma\) consensus sequence were labeled with \([\gamma\text{-}32P}\) ATP using T4 polynucleotide kinase and were purified in Bio-Spin chromatography columns (Bio-Rad, Hercules, CA). Twenty five micrograms of nuclear protein were preincubated with EMSA buffer (12 mM HEPES (pH 7.9), 4 mM Tris-HCl (pH 7.9), 25 mM KCl, 5 mM MgCl2, 1 mM EDTA, 1 mM DTT, 50 ng/ml poly(d(I-C)), 12% glycerol (v/v), and 0.2 mM PMSF) on ice for 10 min before addition of the radiolabeled oligonucleotide for an additional 10 min. Protein-nucleic acid complexes were resolved using a nondenaturing polyacrylamide gel consisting of 5% acrylamide (29/1 ratio of acrylamide/bisacrylamide) and were run in 0.5x TBE (45 mM Tris-HCl, 45 mM boric acid, and 1 mM EDTA) for 1 h at constant current (30 mA). Gels were transferred to Whatman 3M paper (Clifton, NJ), dried under a vacuum at 80°C for 1 h, and exposed to photographic film at -70°C with an intensifying screen. Densitometric analysis was performed using ImageQuant (Molecular Dynamics, Sunnyvale, CA).
NF-κB (p65) Activation

NF-κB (p65) activation was detected in lung nuclear extracts by using an ELISA based transcription factor assay kit (TransAM NF-κB p65; Active Motif, Carlsbad CA) as previously described (6, 26). Briefly 20μg of lung nuclear extract was added to a 96-well plate coated with the NF-κB consensus site (5’ –GGGACTTTCC-3’) and incubated for 1h at room temperature followed by washes and addition of a primary antibody specific for the p65 subunit. Following this, a HRP-conjugated secondary antibody was added and a colorimetric readout measured at 450nM on a spectrophotometer. To monitor the specificity of the assay competition was performed by adding wild type or mutated oligonucleotides at a concentration of 20 pmol per well.

Statistical Analysis

Data was analyzed using SigmaStat for Windows Version 3.10 (SysStat Software, San Jose, CA). MAP values are expressed as mean ± SD; the remainder of the values in the figures and text are expressed as mean ± SE of n observations, where n represents the number of animals in each group. Specifically, MAP results were analyzed using a two way analysis of variance (ANOVA) with Bonferroni Correction. For the remainder of the data analysis a one way ANOVA or a Kruskal Wallis one way ANOVA on ranks was utilized. A value of $p < 0.05$ was considered significant.
RESULTS

*C-peptide ameliorates Hypotension following resuscitation.* There were no significant differences in MAP at baseline amongst rats in the sham group; similarly rats in the HS/R groups had no significant difference in MAP at baseline and during hemorrhage (Fig. 1A, B). Sham rats had no significant difference in MAP throughout the experiment; additionally treatment with C-peptide did not have any effect on MAP in this group (Fig. 1A, 1B). Following resuscitation at 1h there were no differences in MAP between vehicle and C-peptide-treated rats (Fig. 1A). However, at 3h after resuscitation, C-peptide-treated rats had a significantly higher MAP compared with vehicle-treated rats (101.3±25 vs. 71.8±16.7 mmHg, \( p<0.05 \)) (Fig. 1B). Of note, Despite the differences in MAP, both groups undergoing HS/R had similar levels of plasma lactate at baseline, during hemorrhage and following resuscitation. Hence, our data suggest that C-peptide administration at the time of resuscitation and thereafter ameliorated hypotension in rats undergoing hemorrhagic shock.

*C-peptide does not cause Hypoglycemia.* C-peptide has been demonstrated to affect glucose utilization in diabetic patients (23). Hence we measured whole blood glucose levels at baseline, during hemorrhage and following resuscitation. C-peptide treated rats had no difference in blood glucose levels when compared with vehicle-treated rats at baseline and during hemorrhage. At 1h and 3h after resuscitation blood glucose levels were 107±13 and 95±18 mg/dl and in vehicle-treated and 87±3 and 120±15 mg/dl C-peptide treated rats respectively.

*C-peptide Reduces Plasma Cytokine and Chemokine levels.* Plasma CINC-1 levels were measured at 1h and 3h after resuscitation (Fig. 2). Rats undergoing HS/R has a significant increase in CINC-1 level when compared with rats in both the control and sham groups at both time-points. However, in rats undergoing HS/R, C-peptide treatment significantly reduced CINC-1 levels at 1h and 3h when compared to vehicle-treated rats (Fig. 2). Similarly, C-peptide
treatment reduced plasma IL-1, IL-6 and MIP-1α levels at 1h and 3h after resuscitation in rats undergoing HS/R when compared to vehicle-treated rats (Fig 3A, 3B, 3C). Taken together this data demonstrates that C-peptide treatment reduces plasma chemokine and cytokine levels to varying degrees in a time-course manner following HS/R suggesting that C-peptide may blunt the systemic inflammatory state following HS/R.

C-peptide treatment reduced Lung Neutrophil Infiltration and Injury. Lung neutrophil infiltration is associated with development of lung injury and is a common occurrence following hemorrhagic shock. We assessed lung neutrophil infiltration at 1h and 3h following resuscitation using a myeloperoxidase assay. When compared to levels in control and sham rats, rats undergoing HS/R and vehicle-treatment had a significant increase in myeloperoxidase activity (Fig. 4). However, rats undergoing HS/R who received C-peptide had a significant reduction in myeloperoxidase activity when compared to vehicle-treated rats at 1h and 3h following resuscitation (Fig. 4). On histologic examination HS/R vehicle-treated rats had remarkable alteration in lung architecture specifically increased interstitial edema and inflammatory cell infiltration when compared to sham rats (Fig. 5A, 5B, 5C, 5D). These pathologic findings were attenuated in C-peptide-treated rats (Fig. 5E, 5F). Furthermore, we quantified plasma RAGE levels as an objective surrogate of lung injury in our model. HS/R rats undergoing vehicle-treatment had a significant increase in plasma RAGE levels when compared to control rats (Fig.6). In these rats, C-peptide treatment significantly reduced plasma RAGE levels when compared to vehicle treated rats (Fig. 6).

C-peptide Increased PPARγ Activity in the Lung. C-peptide has been demonstrated to activate the transcription factor PPARγ in opossum kidney cells in vitro and in vivo in the lungs of septic mice (4, 34). To discern the anti-inflammatory effect of C-peptide in our model, we assessed PPARγ nuclear expression and DNA binding in rats undergoing HS/R. By western blot analysis nuclear expression of PPARγ was unchanged in C-peptide treated rats when compared
to vehicle-treated animals. However, vehicle-treated rats that underwent HS/R had a reduction in DNA binding when compared to control rats (Fig. 7). C-peptide treatment in the HS/R group significantly increased DNA binding when compared to vehicle-treated animals (Fig. 7).

**C-peptide reduced ERK1/2 phosphorylation in the Lung.** Since, C-peptide has been reported to also modulate ERK1/2; we assessed ERK1/2 activation in lung extracts in rats undergoing HS/R (5, 34). There was negligible phosphorylation of ERK1/2 in control rats; 3h after resuscitation vehicle-treated rats had a significant increase in ERK1/2 phosphorylation (Fig. 8). On the other hand, C-peptide treated rats had a significant reduction in ERK1/2 phosphorylation when compared to vehicle-treated rats 3h after resuscitation (Fig. 8), thus suggesting a reduction in kinase activation.

**C-peptide reduced activation of AP-1, NF-κB in the Lung.** AP-1 and NF-κB are important transcription factors that modulate the inflammatory response following oxidative stress (2, 24). Hence we investigated the effect of C-peptide administration on AP-1 and NF-κB (p65 subunit) activation in the lung in rats undergoing HS/R. Vehicle-treated rats had a significant increase in AP-1 DNA binding following resuscitation when compared to control rats (Fig. 9). C-peptide treatment significantly reduced AP-1 DNA binding following resuscitation compared to vehicle-treated rats (Fig. 9). Similarly, at 1h and 3h following resuscitation vehicle-treated rats had a 4 fold and 1.6 fold increase in p65 activation respectively compared to control rats (Fig. 10). C-peptide treatment reduced p65 activation when compared to vehicle-treated rats at both time points following resuscitation (Fig. 10). Taken together these results indicate that C-peptide administration during resuscitation blunts activation of important pro-inflammatory transcription factors in the lung.
DISCUSSION

Our current work represents the first study to explore the effects of C-peptide on the inflammatory response and lung injury in the setting of hemorrhagic shock and resuscitation. Notably, when administered during resuscitation following hemorrhagic shock, C-peptide reduced the systemic inflammatory response as shown by a reduction in cytokine (IL-1, IL-6) and chemokine (CINC-1, MIP-1α) levels. Furthermore, C-peptide reduced lung damage, neutrophil infiltration and plasma RAGE levels. In the lung, this beneficial effect was associated with inhibition of pro-inflammatory transcription factors AP-1 and NF-κB and activation of the anti-inflammatory transcription factor PPARγ. These anti-inflammatory effects were notable early after resuscitation and were independent of blood glucose levels.

C-peptide treatment in rats undergoing HS/R was associated with an improvement in blood pressure. These rats had a significantly higher MAP at the 3h time-point following resuscitation when compared to vehicle-treated rats. Notably, C-peptide did not increase MAP in the sham group. Hence, it is possible that amelioration of the hypotensive state could be secondary to the blunted systemic inflammatory response seen with C-peptide treatment in rats subjected to HS/R. However, it has also been shown that C-peptide may have cardio-protective effects following ischemia-reperfusion. In this regard Young et al. in an ex vivo model of global myocardial ischemia reperfusion have demonstrated that C-peptide improves left ventricular contractility (37). Although we did not assess myocardial contractility in our study such an effect may have contributed to our results.

Lung injury is one of the commonest consequences of resuscitated hemorrhagic shock and is characterized by a massive infiltration of neutrophils, which are a major cell type involved in exaggerated lung inflammation seen following resuscitation (1). Previous studies have demonstrated that C-peptide treatment is associated with reduction in lung injury scores in a murine model of endotoxic shock (34). Similarly in our study, we observed that C-peptide
treated rats had a reduction in lung interstitial and alveolar edema, alveolar damage and infiltration of inflammatory cells when compared to vehicle treated rats. The present study also shows that C-peptide’s therapeutic effect on lung damage may be a consequence of a reduction in neutrophil recruitment through inhibition of chemokine release, notably CINC-1 and MIP-1α. Other experimental evidence indicates that C-peptide may interfere with neutrophil adhesion and recruitment (29). Scalia et al. demonstrated a reduction in leukocyte rolling and adherence with reduced endothelial cell surface expression of adhesion molecules (P-selectin and ICAM-1) in rat mesenteric vasculature with C-peptide treatment (29). More recently, C-peptide has been shown to reduce endothelial cell expression of VCAM-1 and a reduction in hyperglycemia mediated production of chemokines interleukin-8 (IL-8) and monocyte chemoattractant protein-1 (MCP-1) (25). It is unlikely that this mechanism contributed to the anti-inflammatory effects seen with C-peptide in our work as hyperglycemia was not noted in either vehicle or C-peptide treated rats. However, these data do suggest that C-peptide may act as an important modulator of leukocyte endothelial cell interaction in the lung thereby impacting neutrophil trafficking and preventing neutrophil-induced lung damage.

Hemorrhagic shock and resuscitation is associated with alveolar epithelial cell injury and death (7). These cells serve an important barrier function in the alveolus and play an important role in alveolar fluid clearance and are involved in the pathogenesis of acute lung injury (36). Recent work has suggested the soluble forms of the protein RAGE which is expressed on alveolar epithelial cells may serve as a marker of alveolar epithelial cell injury (33). Furthermore, plasma RAGE levels in trauma patients were helpful in discriminating patients with acute lung injury (17). We utilized plasma RAGE a potential marker of lung injury in our rodent model. In our study C-peptide treated rats undergoing HS/R had a decrease in plasma RAGE levels at 1h and 3h following resuscitation when compared to vehicle-treated rats. This would suggest a reduction in lung injury, possibly alveolar epithelial cell injury. Whether this reduction in plasma RAGE levels is secondary to the effect of C-peptide mediated reduction in neutrophil infiltration
or a direct effect of C-peptide on RAGE signaling and expression cannot be discerned from our work. Taken together with lung histology and neutrophil infiltration, it would appear that C-peptide does reduce lung inflammation and injury following hemorrhagic shock and resuscitation.

It has been shown that oxidative stress during hemorrhagic shock and resuscitation may activate several mitogen activated protein kinases (MAPK) such as extracellular signal-regulated kinase (ERK 1/2), c-Jun N-terminal kinase (JNK) and IκB kinases (IKK), which mediate the transduction of oxidant stress signals to the nuclear transcription factors activator protein (AP)-1 and nuclear factor-κB (NF-κB) (16). Furthermore, recent studies from our laboratory have demonstrated that whereas the pro-inflammatory responses are exaggerated in the lung following hemorrhagic shock, the anti-inflammatory pathway of the nuclear receptor PPARγ is dramatically depressed (40).

PPARγ is a nuclear transcription factor whose activation is well described to be associated with anti-inflammatory effects in systemic inflammatory processes such as septic shock, myocardial ischemia-reperfusion and hemorrhagic shock (39). In our current work, C-peptide administration induced a significant increase in PPARγ DNA binding in the lung when compared to vehicle-treated rats. Interestingly, nuclear expression of PPARγ was not different between C-peptide treated and vehicle-treated rats. The mechanism by which C-peptide activates PPARγ \textit{in vivo} is unclear. \textit{In vitro} studies in proximal renal tubular cells have shown that C-peptide increases PPARγ transcriptional activity in a concentration dependant manner without altering PPARγ protein expression (4). Similarly, \textit{in vivo} C-peptide administration in endotoxemic mice increased PPARγ activation in the lung with a concurrent blunting of the inflammatory response and lung injury (34). Hence, it appears that C-peptide may act as a PPARγ activator in the lung and thereby may dampen the inflammatory response.
NF-κB is a major pro-inflammatory transcription factor that is activated early after hemorrhage and resuscitation and is involved in the pathogenesis of lung inflammation and injury (30). The NF-κB protein consists of two subunits p65 and p50. In the quiescent state these subunits exist complexed with inhibitor κB (IκB) proteins in the cytosol. Classically, the activation sequence for this pathway involves phosphorylation of IκBα by an upstream kinase with subsequent degradation and release of the p65 and p50 subunits (38). These subunits translocate to the nucleus where they orchestrate the expression of numerous genes involved in the inflammatory process (38). We assessed activation of the p65 subunit of NF-κB by utilizing a transcription factor ELISA. Following hemorrhage and resuscitation vehicle-treated rats had a significant increase in p65 activation compared to sham rats. C-peptide treatment significantly reduced p65 activation to basal levels compared to vehicle-treated rats. Similarly to our findings, in vitro work in human aortic endothelial cells exposed to hyperglycemia demonstrated an increase in nuclear translocation of the p65 subunit; C-peptide treatment significantly reduced translocation of p65 and binding activity of p50 (25). These results suggest that C-peptide has the ability to inhibit activation of NF-κB subunits by inhibiting their nuclear translocation.

AP-1 represents another important transcriptional factor that regulates the synthesis of a multitude of pro-inflammatory proteins (24). Vehicle-treated rats had a significant increase in AP-1 DNA binding following resuscitation when compared to sham rats. C-peptide treatment significantly reduced AP-1 DNA binding following resuscitation compared to vehicle-treated rats. In support of these findings, we have also observed that AP-1 activation is inhibited in the kidney following resuscitation with C-peptide treatment and this event is associated with improved kidney function (13). The mechanism by which C-peptide may effect NF-κB and AP-1 activation is unclear. Our study suggests that C-peptide may modulate the upstream activation of MAPK signaling. C-peptide has been previously described to modulate the activation of
ERK1/2 members of the MAPK family that regulates the inflammatory response (18). For example, C-peptide treatment reduced ERK1/2 in the lung in endotoxemic mice (34). On the contrary, in vitro studies have demonstrated that C-peptide may activate ERK1/2 as shown by Al-Rasheed et al. in opossum kidney cells thereby impacting cell proliferation (5). It has also been reported that following trauma-hemorrhage ERK1/2 activation is associated with the development of a pro-inflammatory state that may contribute to organ injury (19). In support of a pathogenic role of these kinases, experimental studies have shown that ERK1/2 inhibition may be beneficial in reducing organ injury following trauma hemorrhage (3, 21). In our study, vehicle-treated rats had a significant activation of ERK1/2 in lung extracts as compared to sham rats. C-peptide treatment reduced ERK1/2 phosphorylation when compared to vehicle-treated rats. Taken together, these data suggest that in vivo C-peptide may act as a potent inhibitor of ERK1/2 activation. Other studies from our laboratory also demonstrate that C-peptide may inhibit JNK activation in the rodent kidney following hemorrhagic shock (13). Furthermore, in vitro studies suggest the existence of an interesting cross-talk between kinases such as ERK1/2 and PPARγ. For example, it has been shown that ERK1/2-mediated serine phosphorylation is associated with a reduction in PPARγ translational activity in placental and kidney cell lines (3, 10). Furthermore, PPARγ activation is associated with inhibition of ERK1/2 in vascular smooth muscle cells (8); on the contrary, ERK1/2 inhibition has been shown to reactivate PPARγ in adipocytes exposed to cyclic stretch (32). Thus, following hemorrhagic shock and resuscitation C-peptide may impact cross talk between ERK1/2 and PPARγ resulting in enhanced anti-inflammatory effects in the lung.

Our work has limitations. The nature of the cell type being affected by C-peptide in our model is unclear. It appears that C-peptide may exert its effects by interacting with a receptor on the cell surface (28). However, despite some work suggesting that this may be a G-protein coupled receptor, the nature of its receptor remains unclear (35). Additionally, it is unclear
whether the effects on cell signaling represent direct or indirect actions of this peptide. Further work will need to be performed to elucidate the mechanism by which C-peptide modulates cell signaling. Additionally, the effects of C-peptide on specific cell types in the lung will need to be defined.

In conclusion, our study represents the first to explore the effects of C-peptide in an *in vivo* model of hemorrhagic shock. C-peptide administered following hemorrhagic shock was associated with blunting of the systemic inflammatory response and a reduction in lung inflammation. In the lung these effects were associated with PPARγ activation and ERK1/2, NF-κB and AP-1 inhibition. These findings are novel and clinically relevant as C-peptide was administered during resuscitation. Hence, C-peptide may represent a novel endogenous peptide with anti-inflammatory properties.
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REFERENCES

FIGURE LEGENDS

Figure 1.
Effect of in vivo treatment with vehicle or C-peptide on mean arterial pressure (MAP) (A) Time course depiction of MAP up to 1h following resuscitation. (B) Time course depiction of MAP up to 3h following resuscitation. Each data point represents the mean ± SD 3 rats in sham groups and 5 rats in HS/R groups. *p < 0.05 vs. HS/R+Vehicle group.

Figure 2.
Effect of in vivo treatment with vehicle or C-peptide on plasma CINC-1 levels at 1h and 3h following HS/R. Each data point represents the mean ± SE of 3 rats in sham groups, 5 rats in control and HS/R groups. *p < 0.05 vs. control group; # p < 0.05 vs. HS/R+ Vehicle group.

Figure 3.
Effect of in vivo treatment with vehicle or C-peptide on plasma levels of IL-1 (A), IL-6 (B), MIP-1α (C) at 1h and 3h following HS/R. Each data point represents the mean ± SE of 5 rats. *p < 0.05 vs. control group; # p < 0.05 vs. HS/R+ Vehicle group.

Figure 4.
Effect of in vivo treatment with vehicle or C-peptide on lung myeloperoxidase activity at 1h and 3h following HS/R. Each data point represents the mean ± SE of 3 rats in sham groups, 5 rats in basal and HS/R groups. *p < 0.05 vs. control group; # p < 0.05 vs. HS/R+ Vehicle group.

Figure 5.
Effect of in vivo treatment with vehicle or C-peptide on lung histology at 3h after HS/R. (A) Representative photomicrograph of histology showing lung architecture from a rat in the Control
group (magnification 100x) (B) Magnified section of prior image (indicated by circle) to show alveolar architecture and blood vessel from a rat in the Control group (magnification 400x). (C) Representative photomicrograph of histology showing lung architecture from a rat in the HS/R+Vehicle group (magnification 100x). (D) Magnified section of prior image (indicated by circle) demonstrating interstitial edema and inflammatory cell infiltration from a rat in the HS/R+Vehicle group (magnification 400x). (E) Representative photomicrograph of histology showing lung architecture from a rat in the HS/R+C-peptide group (magnification 100x). (F) Magnified section of prior image (indicated by circle) demonstrating a reduction in interstitial edema and inflammatory cell infiltration from a rat in the HS/R+C-peptide group (magnification 400x).

Figure 6.

Effect of in vivo treatment with vehicle or C-peptide on plasma RAGE level at 1h and 3h following HS/R. Each data point represents the mean ± SE of 5 rats. *p < 0.05 vs. control group; # p < 0.05 vs. HS/R+Vehicle group.

Figure 7.

Effect of in vivo treatment with vehicle or C-peptide on PPARγ DNA binding in lung at 3h following HS/R. (A), Representative autoradiograph of EMSA for PPARγ; (B), image analysis of PPARγ DNA binding determined by densitometry. Each data point represents the mean ± SE of 5 rats from two separate experiments. *p < 0.05 vs. control group; # p < 0.05 vs. HS/R+Vehicle group.
**Figure 8.**

Effect of *in vivo* treatment with vehicle or C-peptide on phosphorylated form of ERK in lung cytosol extracts at 3h following HS/R. (A), Representative western blot of phosphorylated form of ERK and total ERK. (B), image analysis of nuclear content of phosphorylated form of ERK as determined by densitometry. Each data point represents the mean ± SE of 5 rats from two separate experiments. *p < 0.05 vs. control group; # p < 0.05 vs. HS/R+Vehicle group.

**Figure 9.**

Effect of *in vivo* treatment with vehicle or C-peptide on AP-1 DNA binding in lung at 3h following HS/R. (A), Representative autoradiograph of EMSA for AP-1; (B), image analysis of AP-1 DNA binding determined by densitometry. The fold increase was calculated vs. control value which was set at 1.0. Each data point represents the mean ± SE of 5 rats from two separate experiments. *p < 0.05 vs. control group; # p < 0.05 vs. HS/R+Vehicle group.

**Figure 10.**

Effect of *in vivo* treatment with vehicle or C-peptide on activation of p65 subunit of NF-κB in lung nuclear extracts as determined by transcription factor assay at 1h and 3h following HS/R. The fold increase was calculated vs. control value which was set at 1.0. Each data point represents the mean ± SE of 5 rats. *p < 0.05 vs. control group; # p < 0.05 vs. HS/R+Vehicle group.
Figure 2.
Figure 3.

A.

- Control
- HS/R+Vehicle
- HS/R+C-Peptide

Plasma IL-1 (pg/ml)

Control 1h 3h

B.

- Control
- HS/R+Vehicle
- HS/R+C-Peptide

Plasma IL-6 (pg/ml)

Control 1h 3h

C.

- Control
- HS/R+Vehicle
- HS/R+C-Peptide

Plasma MIP-1α (pg/ml)

Control 1h 3h
Figure 4.
Figure 5.
Figure 6.
Figure 7.

A. PPARγ DNA Binding

B. PPARγ DNA Binding (relative intensity)

- Control
- HS/R+Vehicle
- HS/R+C-Peptide

Bars with * indicate significant difference from Control, and bars with # indicate significant difference from HS/R+Vehicle.
Figure 8.

A. Control  HS/R+Vehicle  HS/R+C-Peptide

B. pERK expression (relative intensity)

- Control
- HS/R+Vehicle
- HS/R+C-Peptide

* #
Figure 9.

A. Control  HS/R+Vehicle  HS/R+C-Peptide

AP-1 DNA Binding

B.

![Bar graph showing AP-1 DNA Binding](image)

- Control
- HS/R+Vehicle
- HS/R+C-Peptide

* and # indicate significant differences.
Figure 10.